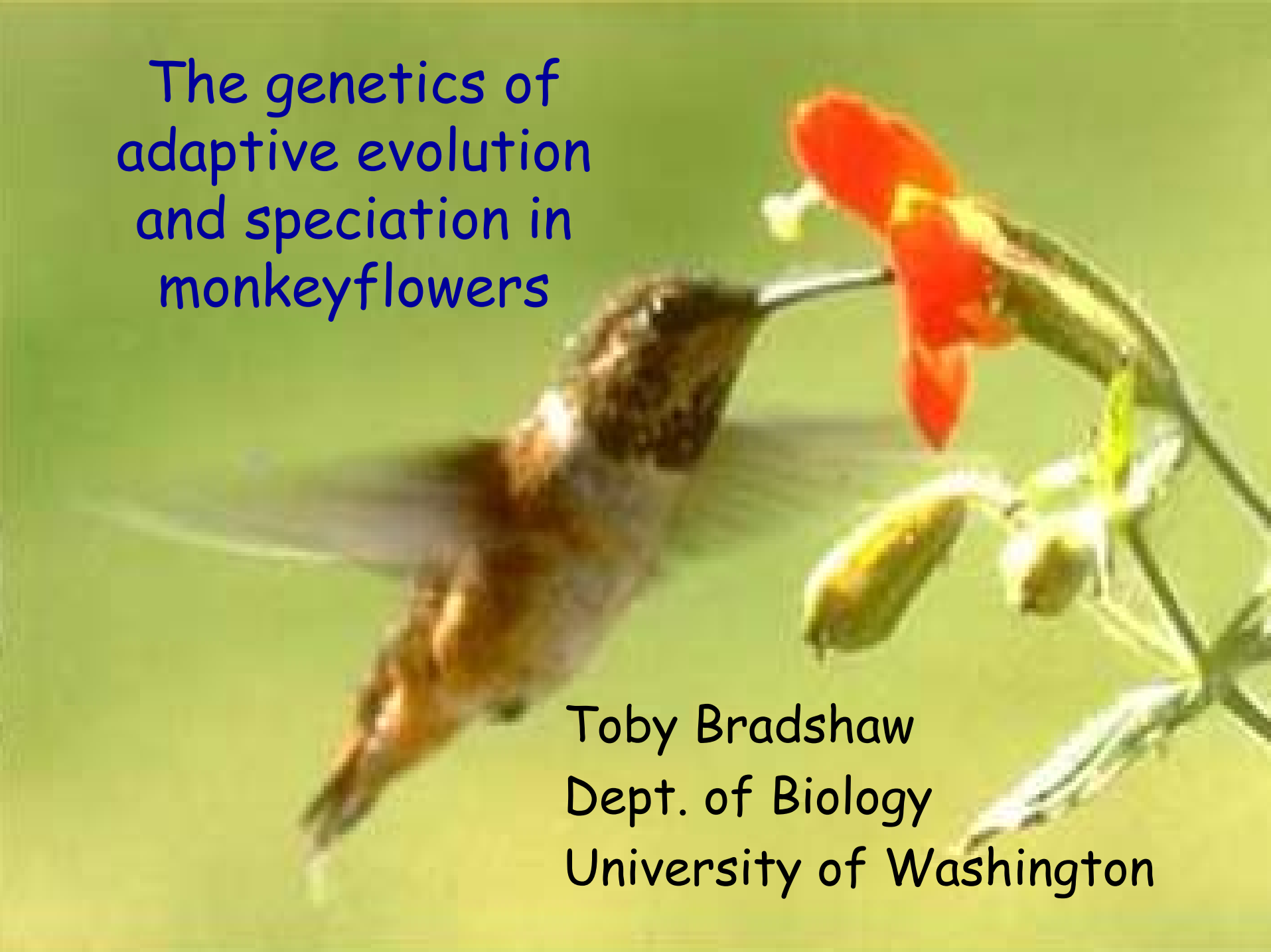


The genetics of
adaptive evolution
and speciation in
monkeyflowers

A photograph of a hummingbird hovering near a red monkeyflower. The bird is in the foreground, slightly out of focus, with its wings spread. The flower is in the background, in focus, showing its bright red petals and yellow center. The background is a soft, out-of-focus green field.

Toby Bradshaw
Dept. of Biology
University of Washington

Why are there species?

“... the living world is not a formless mass of randomly combining genes and traits, but a great array of ... gene combinations, which are clustered on a large but finite number of adaptive peaks.” – Theodosius Dobzhansky





Mimulus

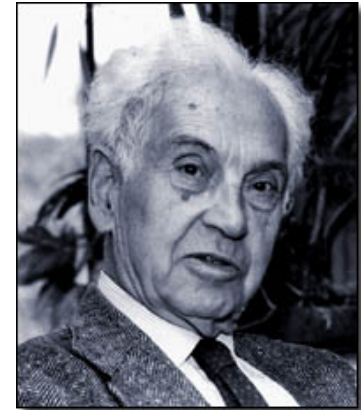
Mimulus as a model for ecological and evolutionary genetics

- ~160 species worldwide, with major radiations in western North America and Australia
- well-resolved phylogeny (except for the *M. guttatus* species complex)
- occupy habitats from sea level to alpine, desert to riparian to aquatic
- life history/habit/mating system diversity – annual, perennial, herbaceous, woody shrub, selfing, outcrossing, mixed, obligately asexual
- phenomenal floral diversity associated with pollination syndromes

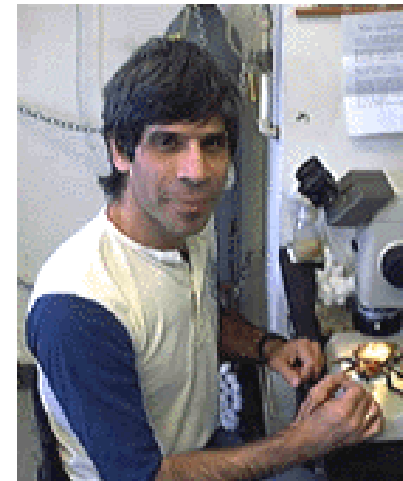
Speciation

Biological species concept

“Groups of actually or potentially interbreeding populations reproductively isolated from all other such groups.” – Ernst Mayr



“When we understand the origin of reproductive isolation, we understand the origin of species.” – Jerry Coyne



Reproductive isolation

Pre-mating barriers to gene flow

- Geographic
- Ecological
- Phenological
- Behavioral
- Mechanical

Post-mating barriers to gene flow

- Gamete incompatibility
- Sperm competition
- Hybrid inviability
- Hybrid sterility
- Hybrid breakdown

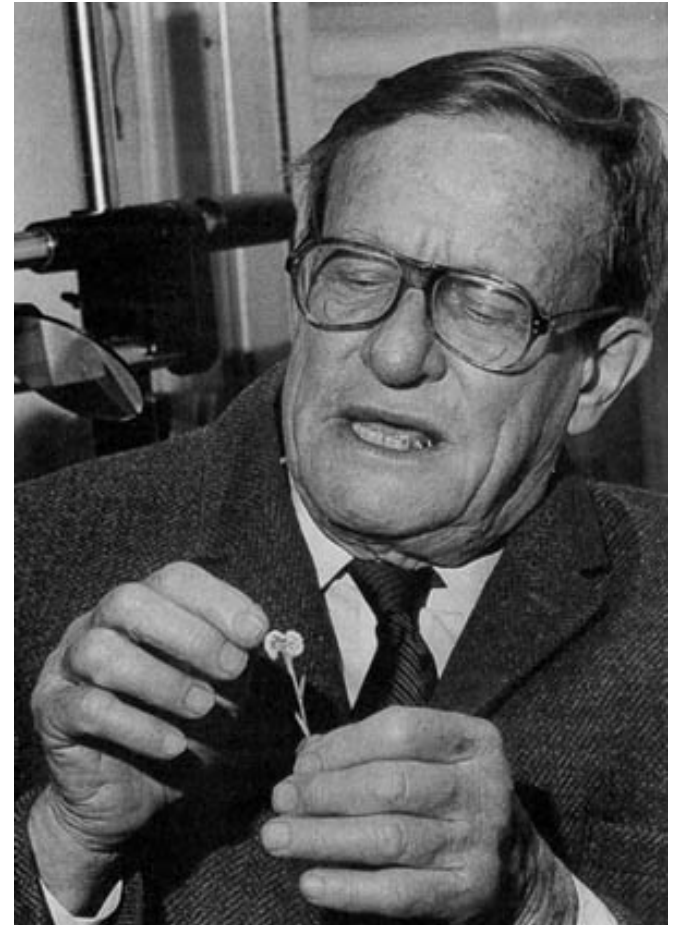
Jordan's law

“Given any species in any region, the nearest related species is not likely to be found in the same region nor in a remote region, but in a neighboring district separated from the first by a barrier of some sort.” -- David Starr Jordan (1905) *Science* 22: 545-562.



The origin of reproductive isolation by ecogeography

“By far the commonest type of isolation in both the plant and the animal kingdom is that resulting from the existence of related types in different geographical regions which differ in climatic and edaphic conditions.” –
G. Ledyard Stebbins (1950)
Variation and Evolution in Plants



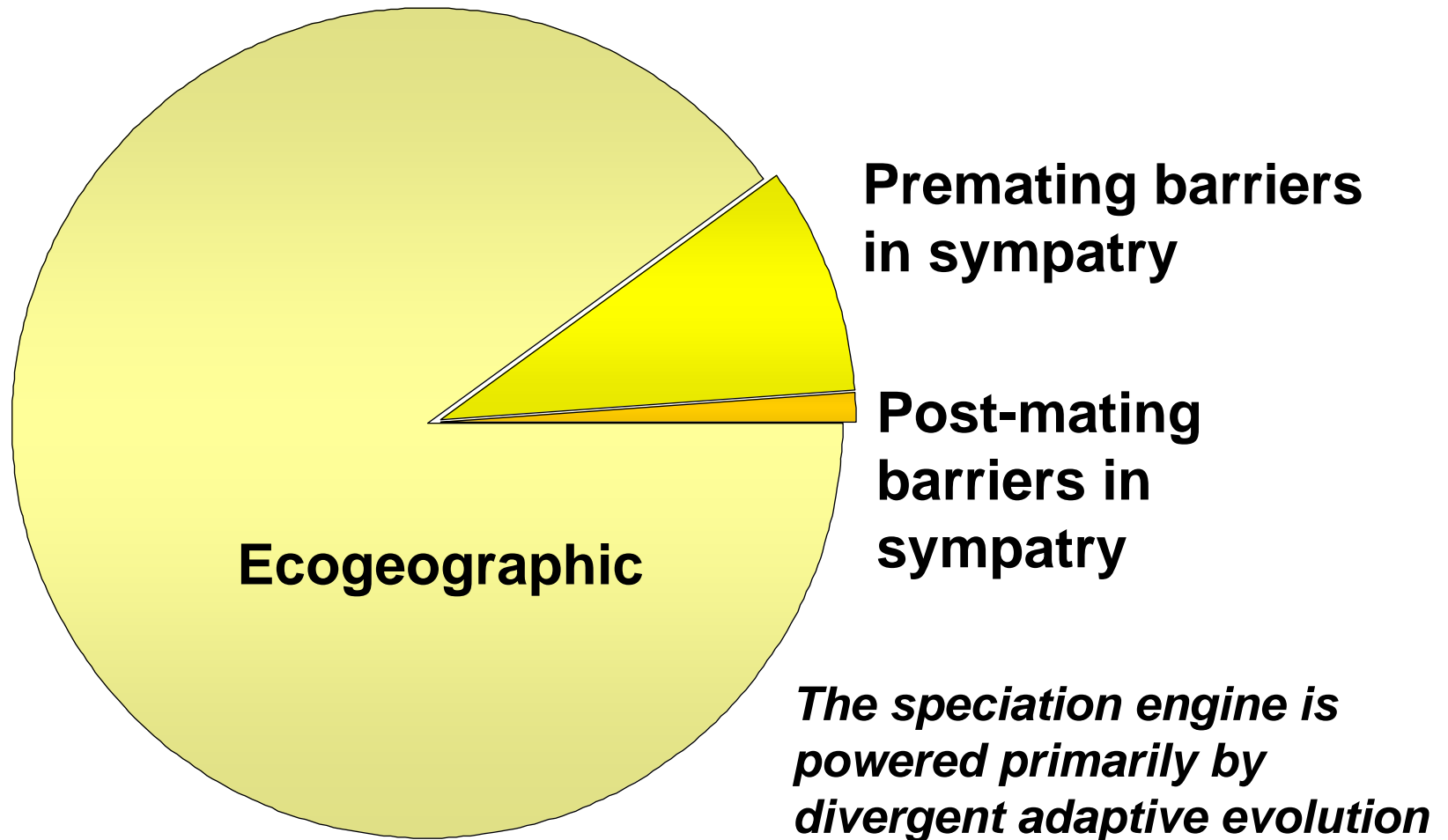
The origin of reproductive isolation by ecogeography

“... not a single geographic race is known that is not also an ecological race; nor is there an ecological race that is not at the same time at least a microgeographic race.”

**-- Ernst Mayr (1963)
*Animal Species and Evolution***



The Jordan/Stebbins/Mayr pie diagram for mechanisms producing reproductive isolation



Why use non-model plant systems?

Good genomics is faster than good ecology

- The whole *Mimulus* genome can be sequenced in less than one greenhouse generation, and in about 10% the length of a field season.

Genetic model systems are usually chosen for the same reasons that ecologists avoid them

- Small size, adapted as human commensals or to human-disturbed habitats

Mimulus as a model for ecological and evolutionary genetics



>70 years of work on ecology, evolution, and genetics, starting with Clausen, Keck, and Hiesey in 1929



M. lewisii



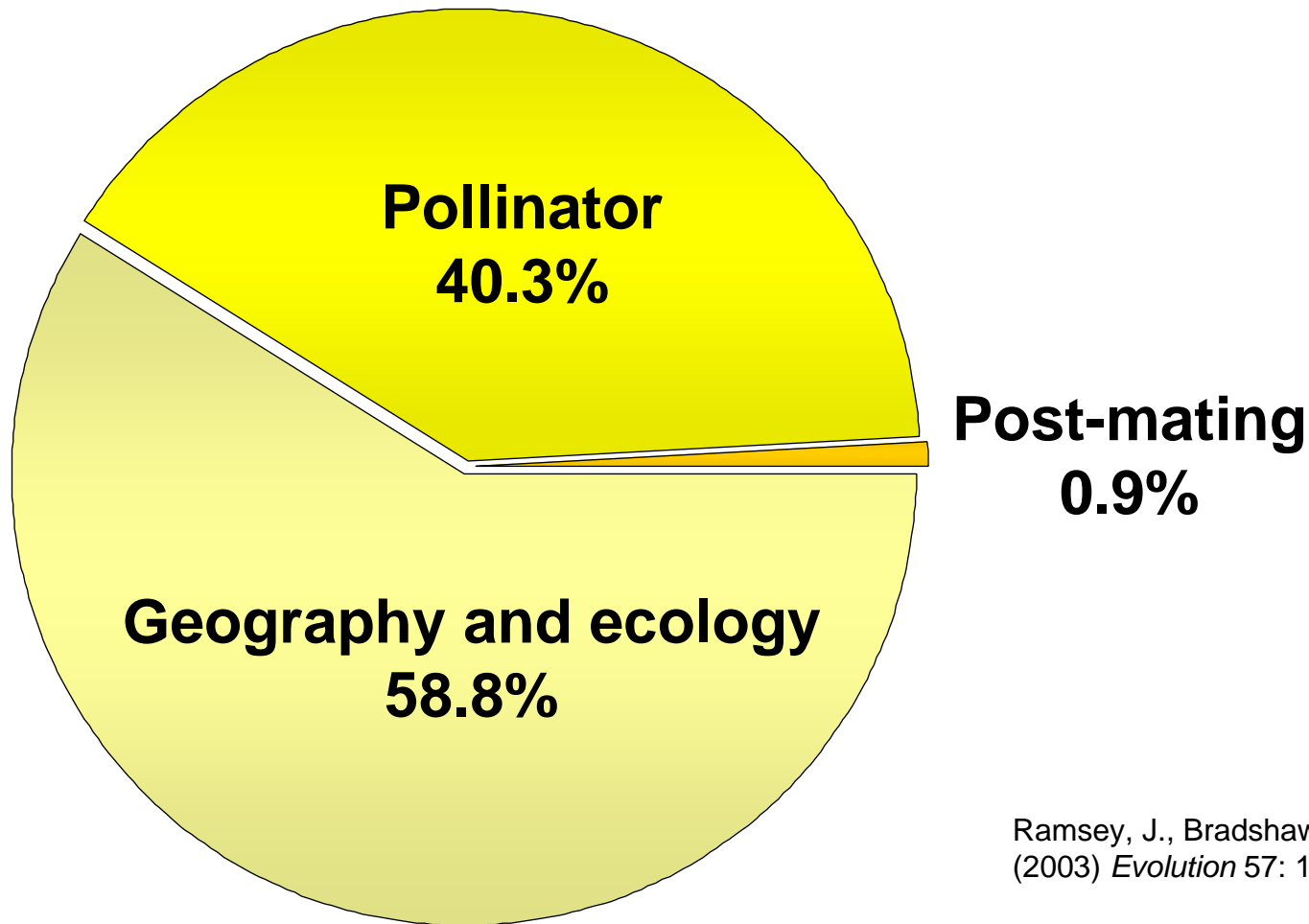
Bumblebee-pollinated
Pink
Wide corolla opening
Inserted stigma/anther
1-2 μ l nectar
Mid-high elevation

M. cardinalis



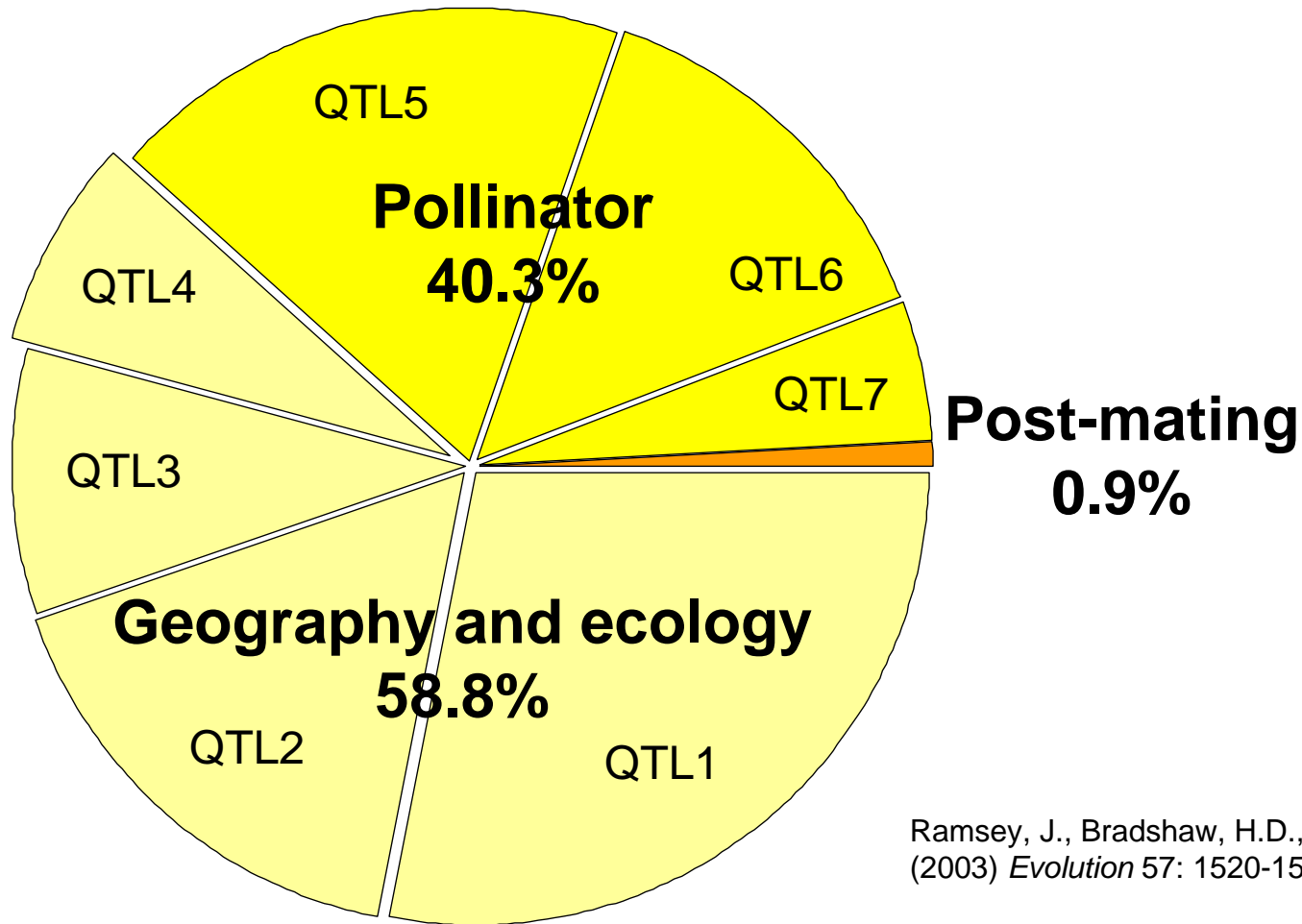
Hummingbird-pollinated
Red
Narrow, tubular corolla
Exserted stigma/anther
40-100 μ l nectar
Low-mid elevation

Components of reproductive isolation between *M. lewisii* and *M. cardinalis*



Ramsey, J., Bradshaw, H.D., Jr., & Schemske, D.W. (2003) *Evolution* 57: 1520-1534.

Components of reproductive isolation between *M. lewisii* and *M. cardinalis*



Ramsey, J., Bradshaw, H.D., Jr., & Schemske, D.W. (2003) *Evolution* 57: 1520-1534.

EXPERIMENTAL STUDIES ON THE
NATURE OF SPECIES

V. BIOSYSTEMATICS, GENETICS, AND
PHYSIOLOGICAL ECOLOGY OF THE ERYTHRANTHE
SECTION OF MIMULUS

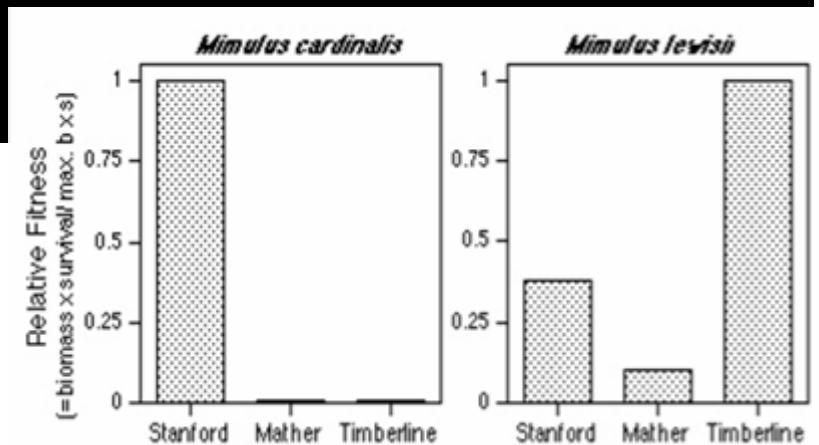
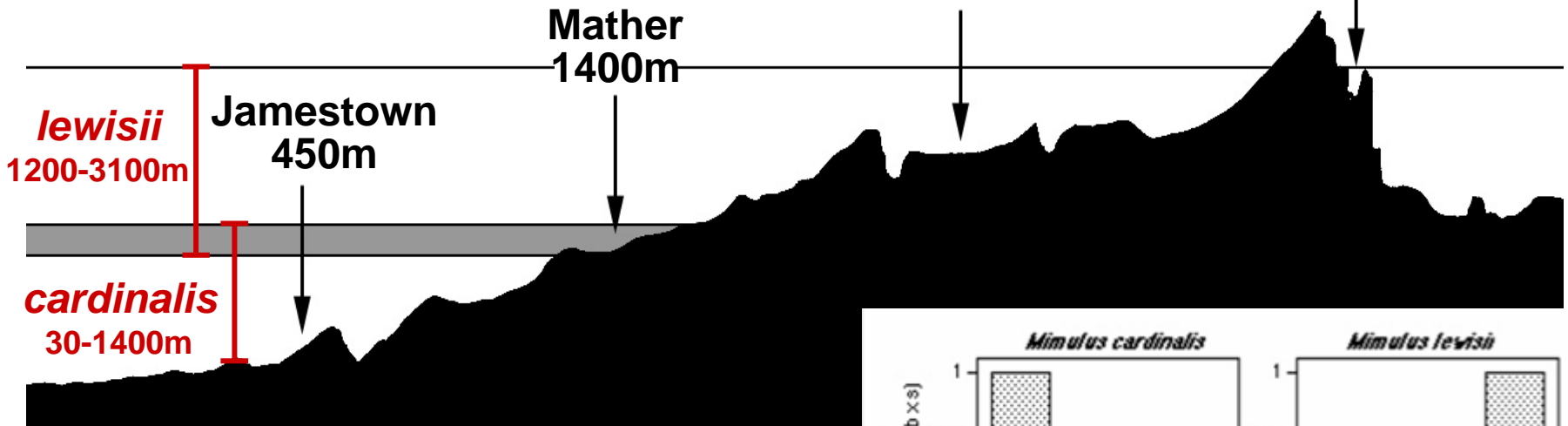
WILLIAM M. HIESEY
MALCOLM A. NOBS
OLLE BJÖRKMAN

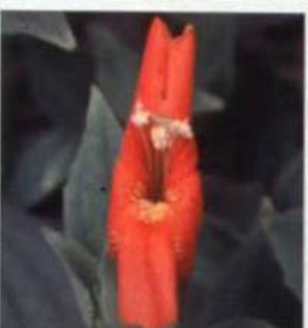


White Wolf
2200m



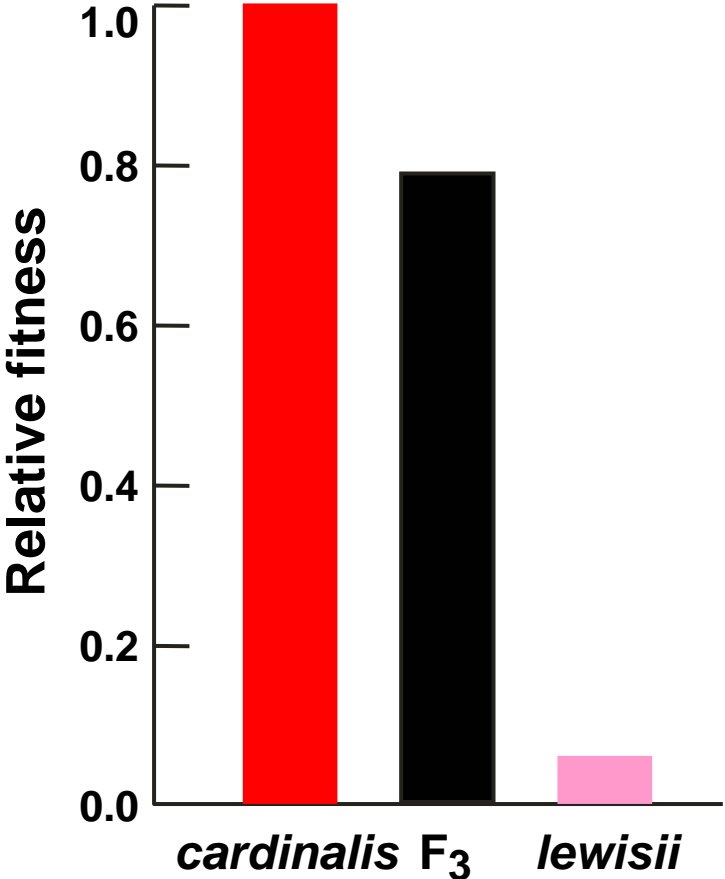
Timberline
3050m



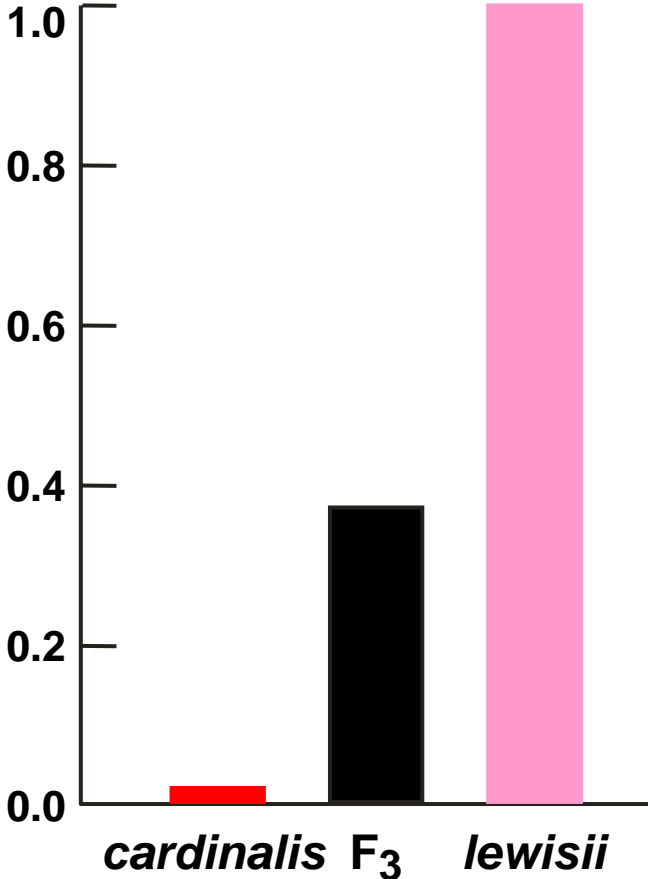


Relative Fitness of Parents and F1 Hybrids

Jamestown
(lo = *cardinalis* habitat)



White Wolf
(hi = *lewisii* habitat)



Experimental evolution in nature: the “ Δp ” Experiment



M. cardinalis
30m-1400m

X

M. lewisii
1200m-3100m



F₁

F₂

F₃

field transplant
N = 4,755 seedlings

field transplant
N = 8,032 seedlings

low elevation
(Jamestown,
450m)

greenhouse
N = 500
seedlings

high elevation
(White Wolf,
2200m)

fates

dead

survive
to flower

survive
to flower

survive
to flower

dead

genotype
random sample with
genome-wide markers

low elevation
survivors

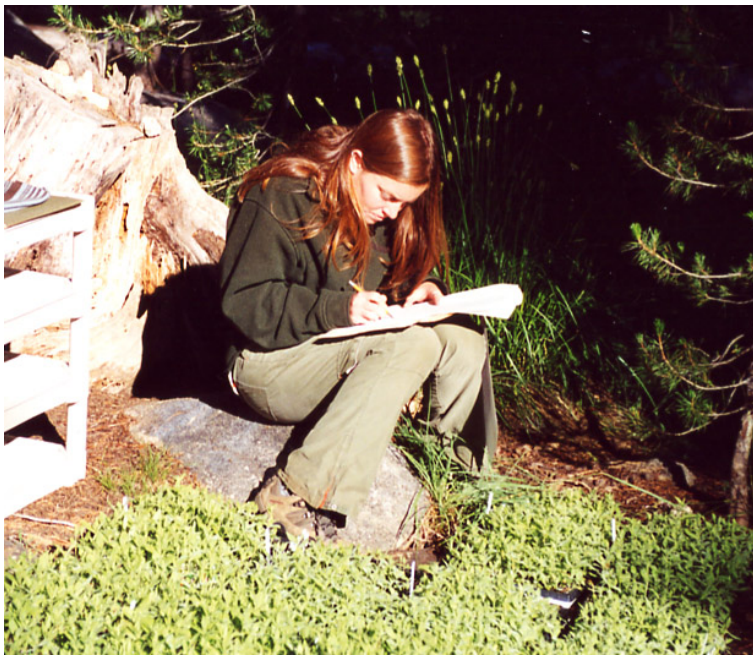
greenhouse
survivors

high elevation
survivors

calculate the change in allele
frequency at each locus
across the genome

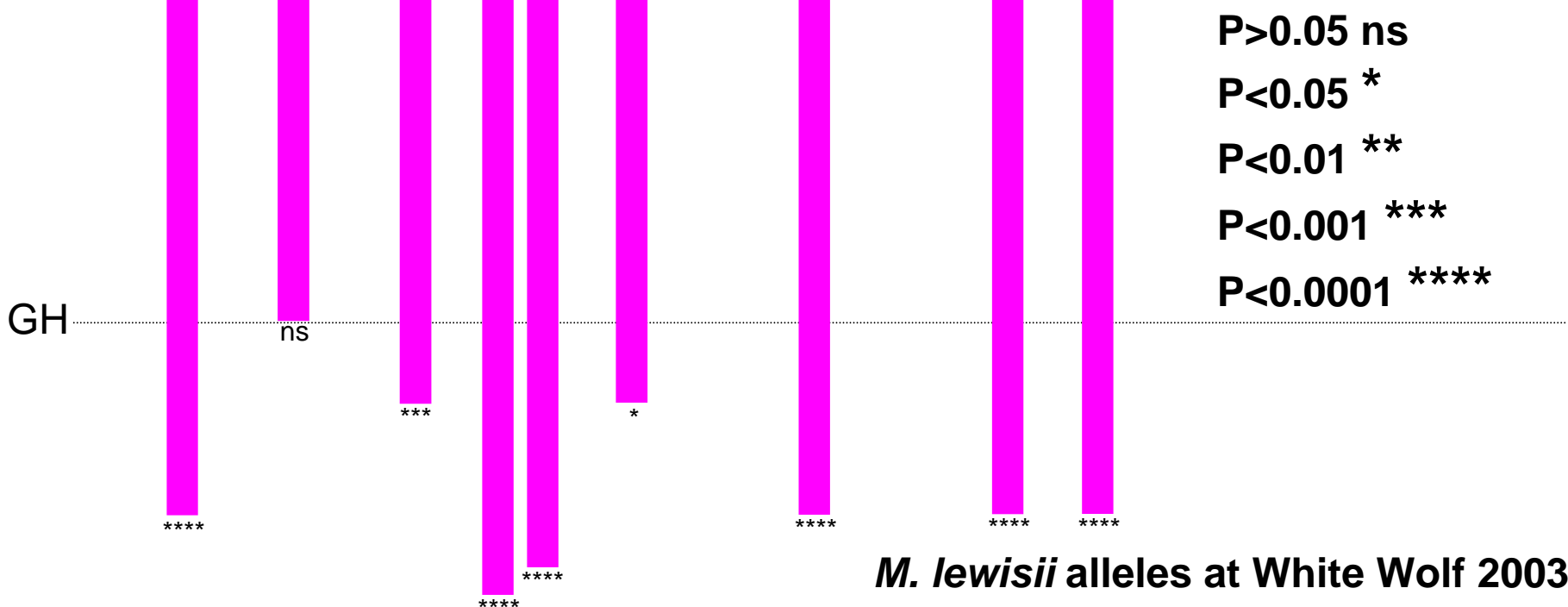
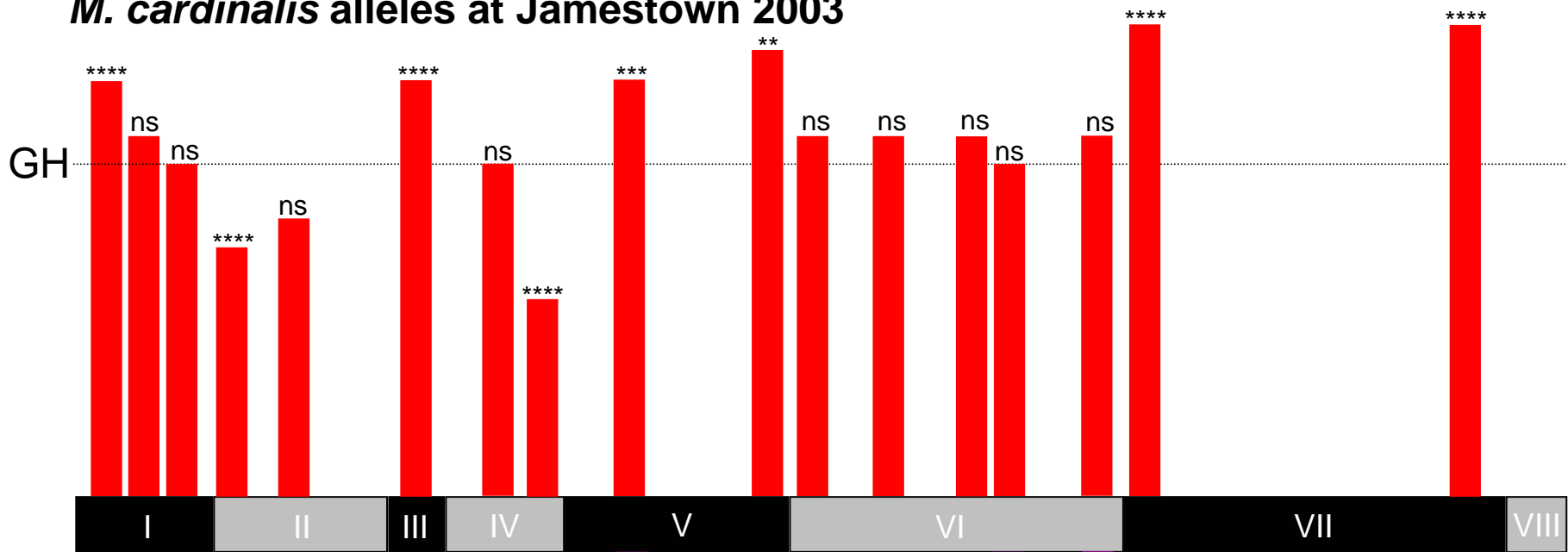
Δp low
elevation

Δp high
elevation





M. cardinalis alleles at Jamestown 2003

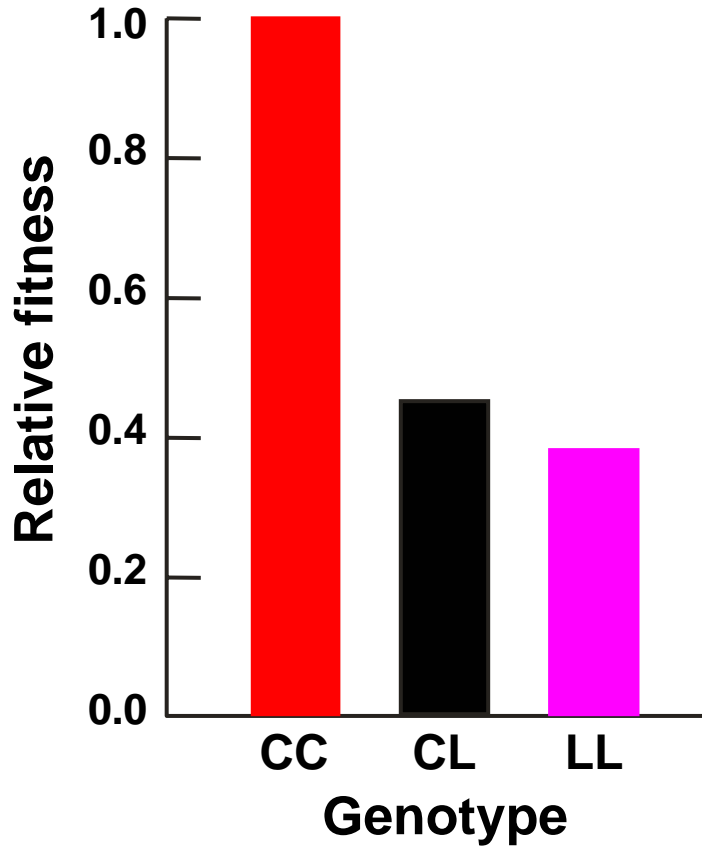


P>0.05 ns
 P<0.05 *
 P<0.01 **
 P<0.001 ***
 P<0.0001 ****

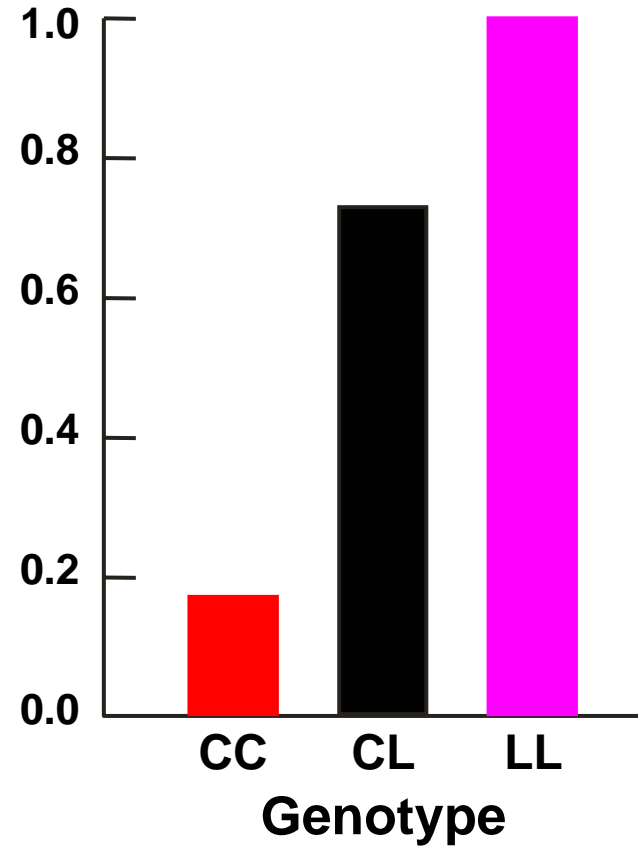
M. lewisii alleles at White Wolf 2003

Relative fitness of F₃ genotypes at MgSTS46 (LG III)

Jamestown
(lo = *cardinalis* habitat)



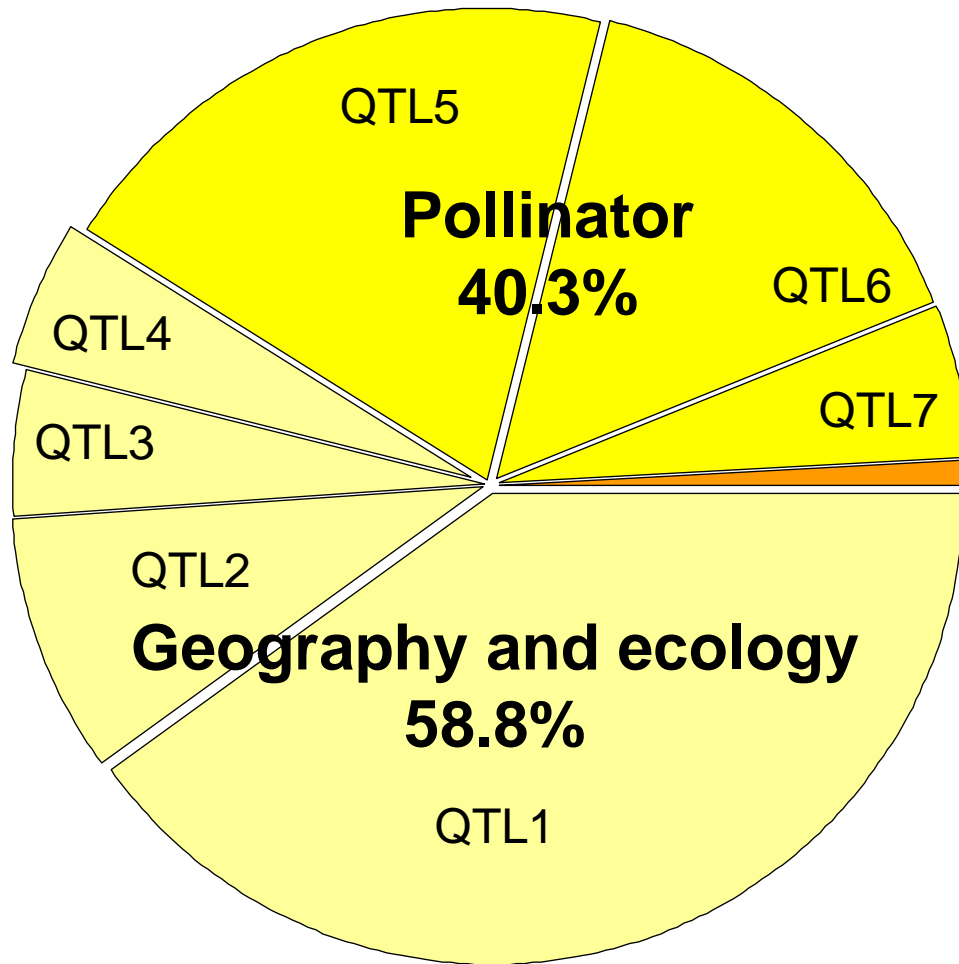
White Wolf
(hi = *lewisii* habitat)



Conclusions

- Differential adaptation is responsible for most of the reproductive isolation between *M. lewisii* and *M. cardinalis* (and, according to Jordan's law, most other pairs of sister taxa).
- QTLs distributed throughout the genome affect local adaptation, with some showing fitness tradeoffs between environments.

Components of reproductive isolation between *M. lewisii* and *M. cardinalis*

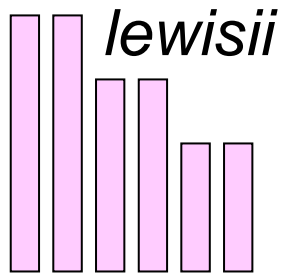


**Post-mating
0.9%**

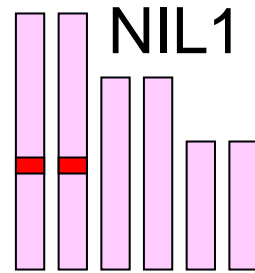
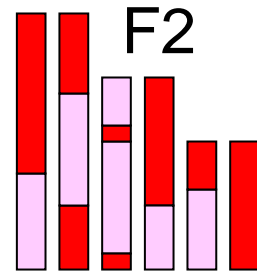
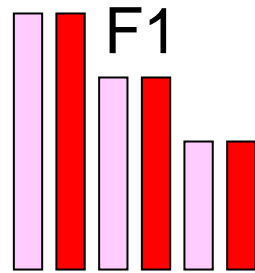
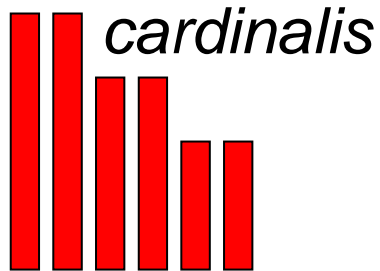


Ramsey, J., Bradshaw, H.D., Jr., & Schemske, D.W. (2003) *Evolution* 57: 1520-1534.

Near-isogenic lines (NILs)



×





Budget

Truck Rental

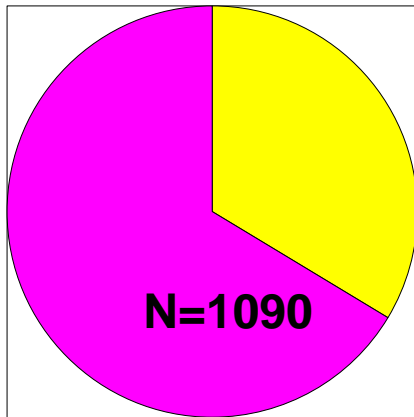
Grumman

7470925

NEWS

1200 347

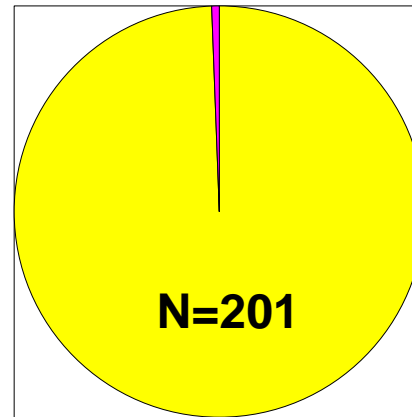




yup
YUP

N=1090

Bumblebees

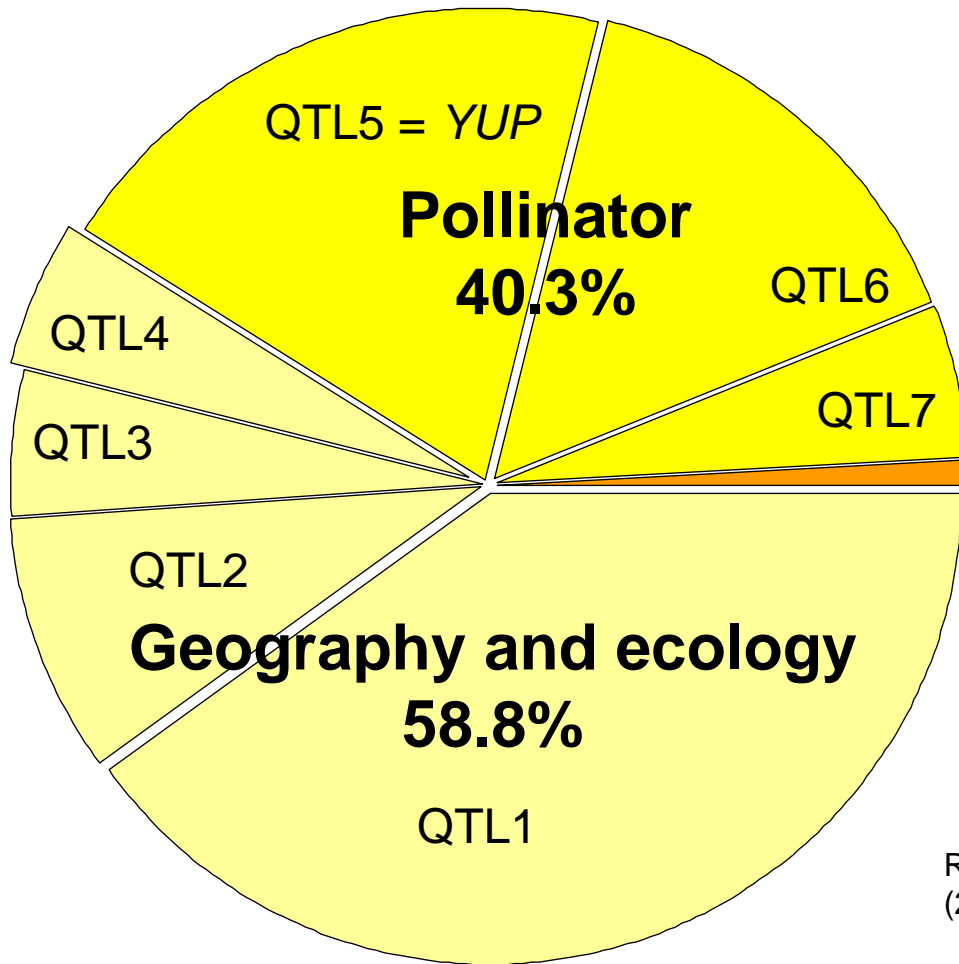


yup
YUP

N=201

Hummingbirds

Components of reproductive isolation between *M. lewisii* and *M. cardinalis*



Post-mating
0.9%



Ramsey, J., Bradshaw, H.D., Jr., & Schemske, D.W.
(2003) *Evolution* 57: 1520-1534.

Conclusions

Major QTLs control all aspects of reproductive isolation in *Mimulus*, from ecogeographic (allopatric) to pollinator preference (sympatric).

The tempo of speciation depends on the genetic details – are these major QTLs single genes, or clusters of linked genes?

From QTLs to genes?

For the traits producing the greatest reproductive isolation in allopatry (**adaptation to elevation**) and sympatry (**carotenoid pigment deposition**) there are no obvious candidate genes. Until there are ...

Two options:

- Positional cloning of major QTLs
- Mutagenesis (at least when one QTL allele is recessive)

Mimulus as a model for ecological and evolutionary genomics

- NSF FIBR project (2004-2009)
- develop genomics tools and apply them to cloning genes involved in speciation
- attract investigators from “traditional” genetic model systems
- recombinant inbred lines (RILs) for mapping
- near-isogenic lines (NILs) for major QTLs
- sub-cM genetic maps based on EST-derived markers
- physical maps from BAC libraries (2006)
- *Mimulus guttatus* genome sequence from JGI in 2007!
- transgenesis
- mutagenesis



LIGHT AREAS

plus:

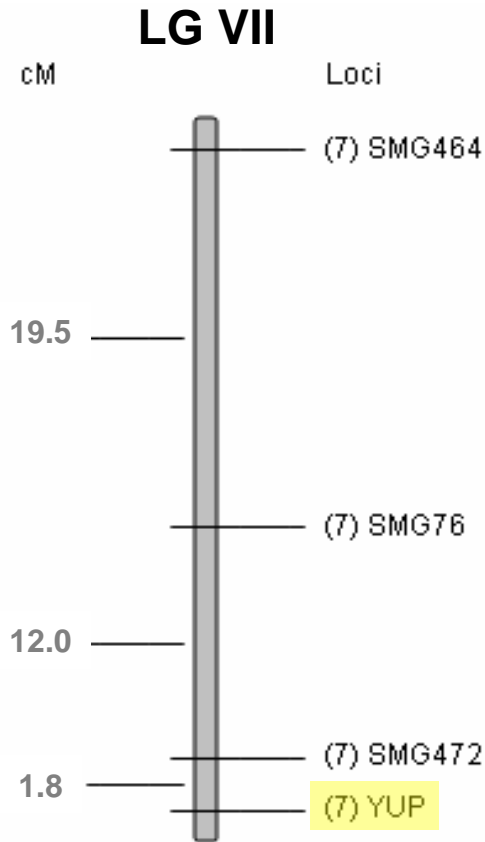
FLOWERING TIME1

PISTIL LENGTH1

~100 new markers from JGI



wt *M. lewisii*



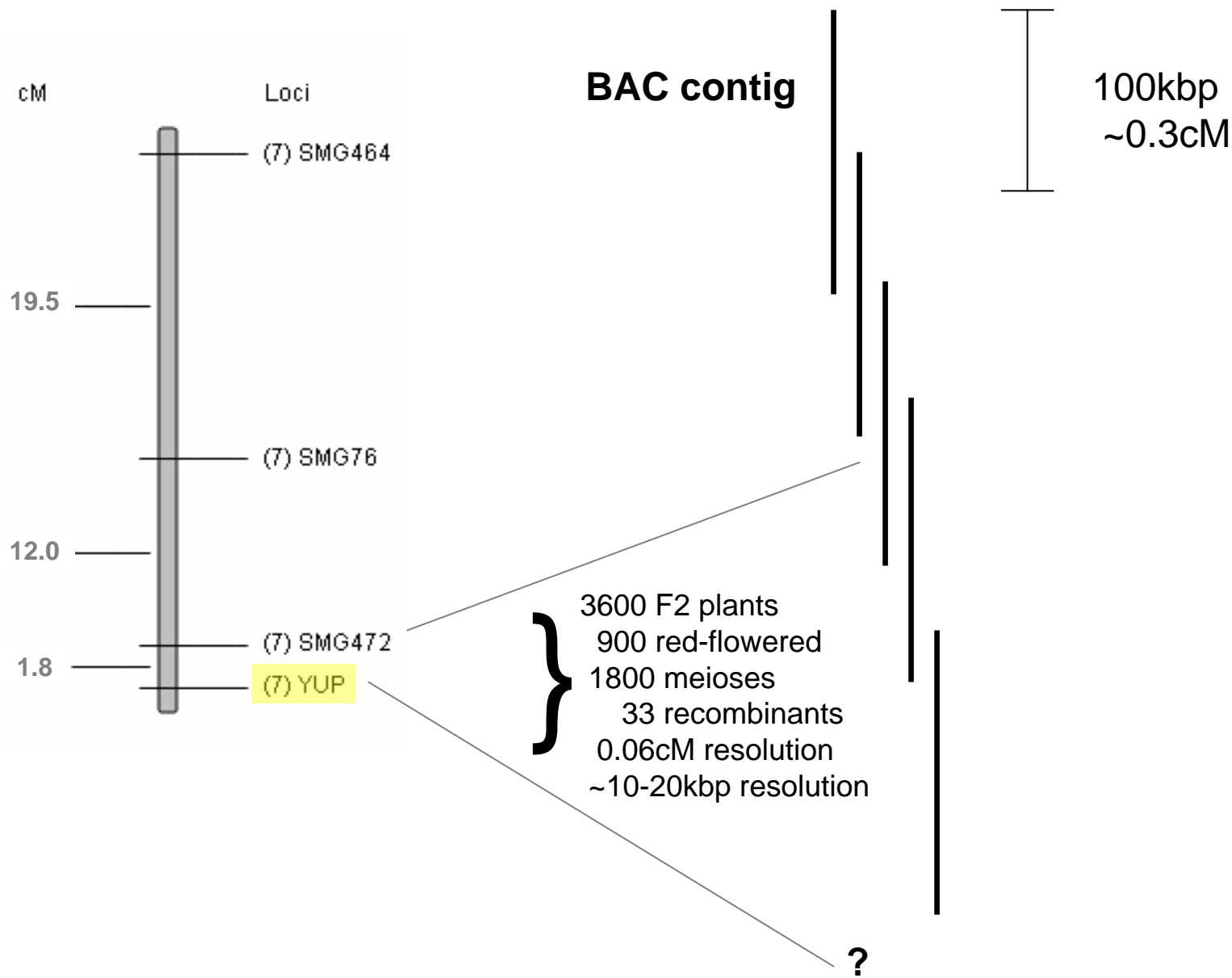
ROSE INTENSITY



REFLEX1

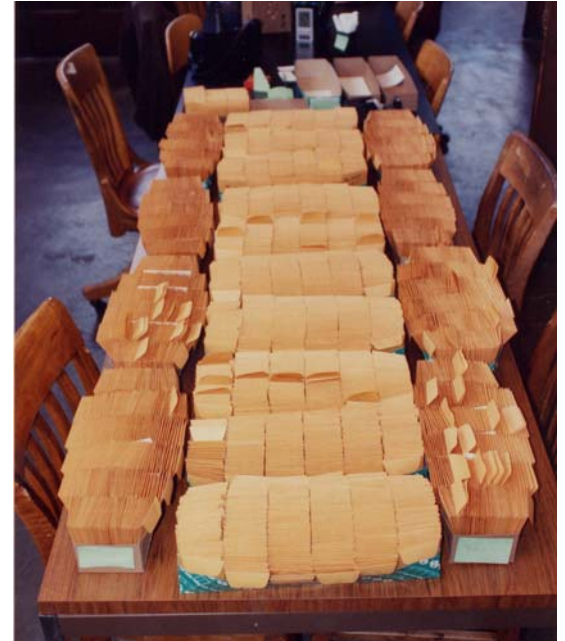


YELLOW UPPER



EMS mutagenesis of *M. lewisii*

- Christina Pince
- 1776 M1 lines
- 31,968 M2 plants



M. lewisii mutants



M. bifidus



M. parishii



M. inconspicuus



wt *M. lewisii*



M. douglasii



M. cardinalis

Conclusions

Many phenotypes that characterize the radiation of floral form across the genus *Mimulus* seem to be controlled by single loci

There is an opportunity to describe a large-scale adaptive radiation at the level of individual genes





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