

Supporting Information

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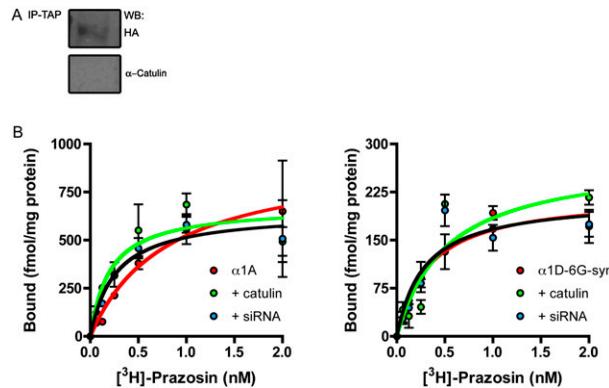


Fig. S1. (A) TAP- α 1A-AR was immunoprecipitated from HEK293 cells, run on SDS/PAGE, and probed for α 1A-AR (HA-tag) and α -catulin. (B) Effects of α -catulin overexpression (green trace) and siRNA knockdown (black trace) were compared with receptor alone (red trace) for both the α 1A-AR (Left) and α 1D-AR (Right). TAP, tandem-affinity purification; AR, adrenergic receptor.

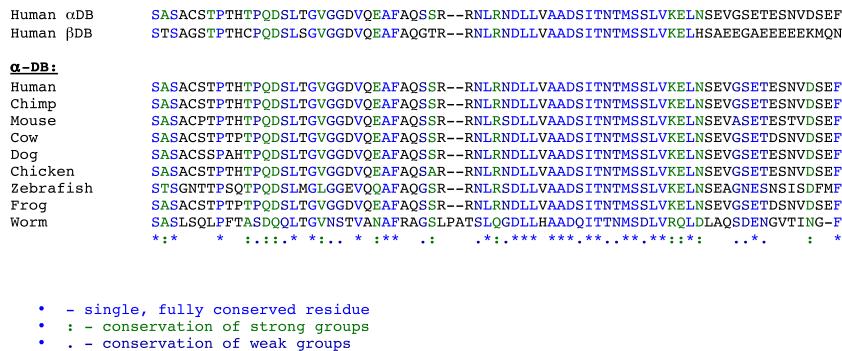


Fig. S2. The C terminus tail of human α -DB1 (amino acids 578–651) was aligned against the C terminus of human β -DB (*Upper*) and the C terminus tails of α -DB genes across multiple species (*Lower*). DB, dystrobrevin.

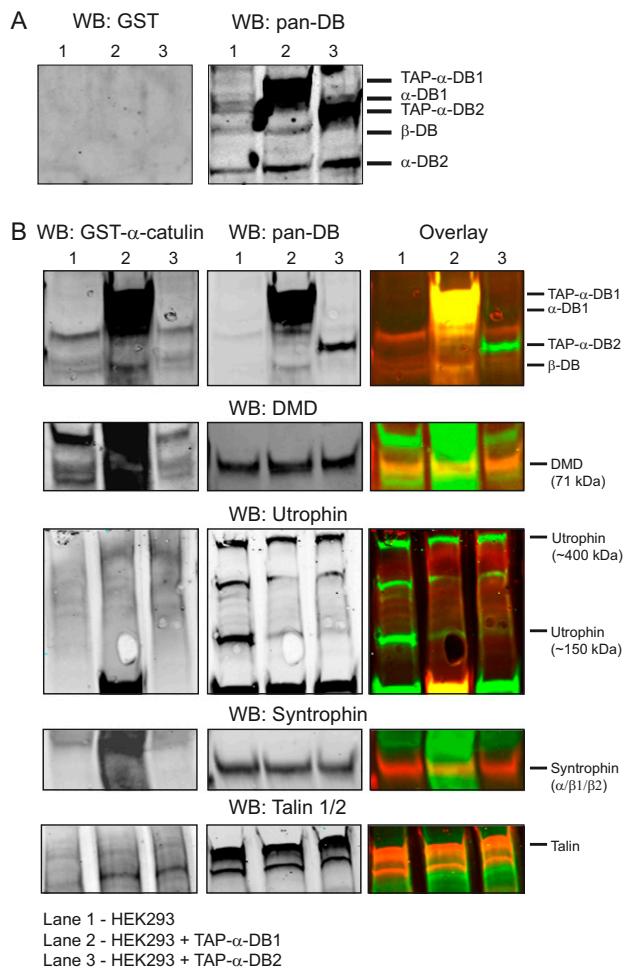


Fig. S3. Untransfected (lane 1), TAP- α -DB1-expressing (lane 2), and TAP- α -DB2-expressing (lane 3) cell lysates were run on SDS/PAGE gels, transferred to nitrocellulose, and overlaid with either (A) GST or (B) GST- α -catulin. The GST Western blot on the right indicates which protein bands GST or GST- α -catulin bound to, center Western blots are as indicated, and left images are the combined images. For dystrophin, syntrophin, and talin combined images, GST- α -catulin is in green and unknowns are in red. For all remaining images, GST- α -catulin is in red and unknowns are in green. TAP, tandem-affinity purification; DB, dystrobrevin.

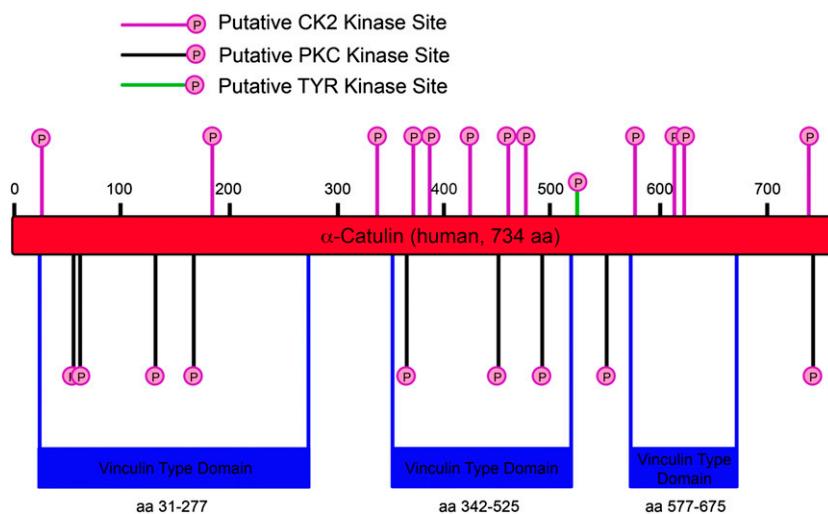


Fig. S4. A pfam protein domain analysis indicated that α -catalin has three domains homologous to vinculin (domains are highlighted in blue). A PROSEARCH motif search found consensus caseine kinase (CK2, pink), protein kinase C (PKC, black), and receptor tyrosine kinase (TYR, green) sites in α -catalin. Approximate locations of phosphorylation sites are indicated.

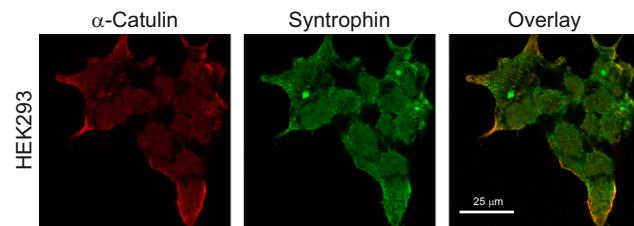


Fig. S5. Syntrophin (green) and α -catulin (red) colocalize in HEK293 cells.

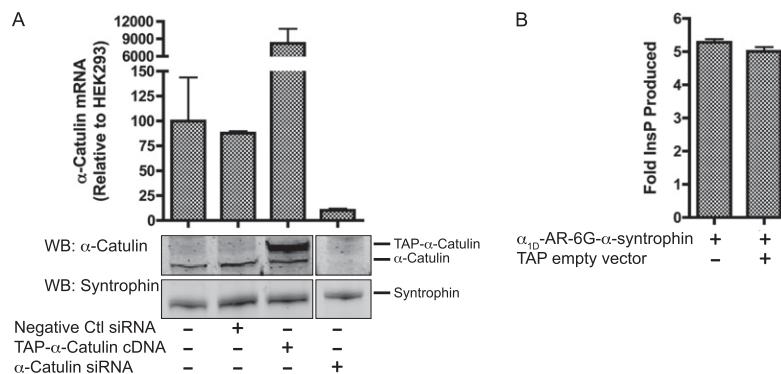


Fig. S6. (A) α -Catulin mRNA and protein levels are measured after treatment for 48 h with siRNA directed against α -catulin. (B) An empty TAP vector was cotransfected with α 1D-AR-6G- α -syntrophin to assess the affect of the TAP vector on α 1D-AR-6G- α -syntrophins IP3 production in response to 100 μ M phenylephrine. Data are graphed as fold over basal measurements. TAP, tandem-affinity purification.

Table S1. TAP/MS results for bait proteins

Bait	DMD		UTR		SNTA1		SNTB1		SNTB2		DTNA		DTNB		CTNNAL1		PPFIA1	
	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%
α_{1D} -AR	—	—	3	2	—	—	10	22	6	12	2	4	—	—	1	3	—	—
α -Syntrophin	34	26	220	41	73	28	2	6	16	29	44	43	9	19	18	24	—	—
β_1 -Syntrophin	10	3	49	17	1	5	7	15	—	—	8	12	—	—	2	2	—	—
β_2 -Syntrophin	21	4	105	34	—	—	—	—	15	22	28	35	1	1	15	20	—	—
α -DB1	7	2	12	5	28	53	63	64	41	43	158	38	—	—	60	44	14	17
α -DB2	8	3	18	7	28	51	64	61	56	48	110	44	—	—	—	—	6	5

Total spectral counts (n) and percentage amino acid coverage of spectral counts are shown for each protein hit. DMD, dystrophin; UTR, utrophin; SNTA1, α -syntrophin; SNTB1, β 1-syntrophin; SNTB2, β 2-syntrophin; DTNA, α -dystrobrevin; DTNB, β -dystrobrevin; CTNNAL1, α -catulin; PPFIA1, liprin- α -1.

Table S2. Complete TAP/MS results for α -catulin

Total	Unique	AA coverage (%)	Best gene name	Description
232	44	60.1	<i>CTNNAL1</i>	α -Catulin
67	30	41.7	<i>DTNA</i>	α -Dystrobrevin
47	41	15.2	<i>UTRN</i>	Utrophin
37	18	31.2	<i>DMD</i>	Dystrophin
27	20	36.5	<i>SNTB2</i>	β 2-Syntrophin
26	17	40.7	<i>SNTB1</i>	β 1-Syntrophin
12	6	17.9	<i>DTNB</i>	β -Dystrobrevin
7	7	8.8	<i>PPFIA1</i>	Liprin- α -1
7	7	10.2	<i>HRNR</i>	Hornerin
5	3	2.8	<i>COL1A1</i>	Collagen α -1(I) chain precursor
4	4	21.6	<i>CAPNS1</i>	Calpain small subunit 1
4	4	12.4	<i>TUBB</i>	Tubulin, β polypeptide
3	3	6.6	<i>HSPA5</i>	HSPA5 protein
3	3	16.0	<i>RPS3</i>	40S ribosomal protein S3
3	3	7.1	<i>SNTA1</i>	α 1-Syntrophin
3	3	11.8	<i>RPS4X</i>	40S Ribosomal protein S4
3	2	1.8	<i>MYLK2</i>	Myosin light chain kinase
2	2	12.1	<i>C1QBP</i>	Complement component 1 Q
2	2	2.4	<i>PPFBP1</i>	Liprin- β -1
2	1	6.8	<i>HBE1</i>	HBE1 protein
2	2	9.6	<i>PPP1CC</i>	Serine/Threonine Protein Phos.
2	1	6.1	<i>SBSN</i>	Suprabasin
1	1	0.8	<i>PLCB2</i>	PLC β 2
1	1	3.4	<i>PLEKHM1</i>	PH domain containing, family M1
1	1	5.5	<i>NDUFA12</i>	13-kDa differentiation-associated protein
1	1	4.6	<i>ITPA</i>	Inosine triphos. pyrophosphatase
1	1	1.9	<i>ALDH5A1</i>	Succinic semialdehyde dehydrogenase
1	1	2.0	<i>SGSH</i>	N-sulphoglucosamine sulphohydrolase
1	1	0.8	<i>ARHGEF5</i>	RhoGEF5
1	1	1.2	<i>EDD1</i>	E3 ubiquitin-protein ligase EDD1
1	1	3.3	<i>TUFM</i>	Tu translation elongation factor
1	1	10.0	<i>PIF</i>	Proteolysis inducing factor
1	1	1.4	<i>MAIL</i>	Nuclear factor of kappa light polypeptide
1	1	9.9	<i>S100A4</i>	Protein S100-A4
1	1	2.6	<i>PLCXD2</i>	Phosphatidylinositol phospholipase CX
1	1	15.6	<i>HEJ1</i>	HEJ1
1	1	5.5	<i>DIRAS2</i>	GTP-binding protein Di-Ras2
1	1	7.4	<i>CALM3</i>	Calmodulin
1	1	8.7	<i>PATE</i>	PATE
1	1	4.6	<i>TUBA1C</i>	Tubulin, α 1C
1	1	3.8	<i>RBMY1D</i>	RNA binding motif protein 2
1	1	0.9	<i>SYCP1</i>	Synaptonemal complex protein 1
1	1	10.8	<i>RPS15A</i>	40S ribosomal protein S15a
1	1	16.3	<i>RPS17</i>	40S ribosomal protein S17
1	1	3.7	<i>GAD1</i>	Putative uncharacterized protein GAD1
1	1	23.9	<i>S100B</i>	Protein S100-B
1	1	1.9	<i>ZHX3</i>	Zinc fingers and homeoboxes protein 3
1	1	0.3	<i>NEB</i>	Nebulin
1	1	0.7	<i>ITIH5</i>	Interalpha inhibitor H5
1	1	1.1	<i>NLRC5</i>	Nucleotide-binding oligomerization domains 27
1	1	0.6	<i>PTPRD</i>	Protein tyrosine phosphatase, receptor type, D
1	1	2.1	<i>IFPS</i>	Ifapsoriasin
1	1	9.7	<i>FLJ46266</i>	Hypothetical protein FLJ46266
1	1	1.3	<i>GTF3C1</i>	General transcription factor IIIC, polypeptide 1
1	1	5.4	<i>C3orf33</i>	Uncharacterized protein C3orf33
1	1	20.0	<i>ZNF14</i>	Zinc finger protein 14
1	1	2.1	<i>KLC3</i>	Kinesin light chain 3
1	1	1.1	<i>XPO5</i>	Exportin-5
1	1	6.2	<i>TUBA8</i>	Tubulin α -8 chain

Total spectral (peptide) counts, unique spectral (peptide) counts, percentage amino acid coverage (AA coverage), gene name (Best gene name), and a brief description of each hit are shown.

Other Supporting Information Files

[Dataset S1 \(XLS\)](#)