



# 10. Interfacing R

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# Interfacing R

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With Bioconductor, R can do a huge proportion of the analyses you'll want – but not everything;

- Intensive (or anachronistic) C++, FORTRAN work, e.g. for pedigrees
- 'Speciality' analyses; some need different computing architecture
- Fancy interactive graphics

However, R can be used to 'manage' other software. In this session, we'll illustrate the basic idea with some favorite examples.

# Starting other software

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NB these commands are for Windows only; see help files for e.g. Unix versions

- `shell()` does the equivalent of a DOS-style command
- `shell("notepad")` starts the Notepad editor
- If the command takes arguments, put them in the same string;  
`shell("notepad myfile.txt")`

The `system()` and `shell.exec()` commands do much the same thing.

# Starting other software

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Some more options for `shell()`;

- `wait`; R 'hangs' until completion
- `translate`; makes forward and backslashes work properly
- `intern`; return the output as an R object

For other options see the `system()` help page, for example `minimized=TRUE`.

Paths for files can be a little messy; `shell()` starts in your working directory (find it using `getwd()`). For files outside of this, give the full pathway.

`paste()` is useful, if you need to do a lot of this sort of thing.

# Examples

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Code for a really mundane job;

```
for(i in 1:100){  
  infile <- paste("gene",i,"data.txt", sep="")  
  outfile <- paste("gene",i,"phase.out", sep="")  
  shell(paste("PHASE",infile,outfile))  
}
```

... this will churn away for hours, although with no error-control.

Why did we use `wait=TRUE` here? (the default)

# Examples

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- WinBUGS implements Bayesian analyses; it's not super-fast but is very flexible
- It needs special (& clever) architecture to achieve this
- WinBUGS' input, output, graphics are all rather clunky
- R is better; so `R2WinBUGS` calls WinBUGS for the difficult bits, and does all the 'translation' itself
- This is done with (repeated) use of `system()`

(Stan has similar functionality, and more state-of-the-art methods for Bayesian calculations.)

# Outline

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Many programs already exist to do useful analyses. It is more convenient to call them from R than to rewrite them in R.

Sometimes this involves calling the C code directly, sometimes just involves using R to write input files for another program

Examples:

- Graphviz: drawing networks
- PMF: input files for ancient Fortran software
- Google Earth: displaying outliers in context.

# Drawing networks

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GraphViz (<http://www.graphviz.org>) is a free program for drawing networks, written by AT&T researchers.

Its input format looks like

```
"15" [shape= box,regular=1 ,height= 0.5 ,width= 0.75 ,style=filled,color= grey ] ;
"16" [shape= circle ,height= 0.5 ,width= 0.75 ,style=filled,color= grey ] ;
"2x3" [shape=diamond,style=filled,label="",height=.1,width=.1] ;
"2"  ->  "2x3"  [dir=none,weight=1]  ;
"3"  ->  "2x3"  [dir=none,weight=1]  ;
"2x3" ->  "1"   [dir=none,weight=2]  ;
"2x3" ->  "4"   [dir=none,weight=2]  ;
"2x3" ->  "5"   [dir=none,weight=2]  ;
"2x3" ->  "6"   [dir=none,weight=2]  ;
```

The `sem` package uses GraphViz to display path diagrams for structural equation models and the `gap` package uses it to draw pedigrees.



# Drawing networks

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In `gap` the `pedtodot()` function writes a GraphViz input file from a pedigree in GAS or LINKAGE format.

	pid	id	fid	mid	sex	aff	GABRB1	D4S1645
1	10081	1	2	3	2	2	7/7	7/10
2	10081	2	0	0	1	1	-/-	-/-
3	10081	3	0	0	2	2	7/9	3/10
4	10081	4	2	3	2	2	7/9	3/7
5	10081	5	2	3	2	1	7/7	7/10
6	10081	6	2	3	1	1	7/7	7/10
7	10081	7	2	3	2	1	7/7	7/10
8	10081	8	0	0	1	1	-/-	-/-
9	10081	9	8	4	1	1	7/9	3/10
10	10081	10	0	0	2	1	-/-	-/-
11	10081	11	2	10	2	1	7/7	7/7
12	10081	12	2	10	2	2	6/7	7/7
13	10081	13	0	0	1	1	-/-	-/-
14	10081	14	13	11	1	1	7/8	7/8
15	10081	15	0	0	1	1	-/-	-/-
16	10081	16	15	12	2	1	6/6	7/7

# Drawing networks

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First the code prints nodes for each individual, with sex and affectedness information

```
for (s in 1:n) cat(paste("\", id.j[s], "\" [shape=",  
  sep = ""), shape.j[s], ",height=", height, ",width=",  
  width, ",style=filled,color=", shade.j[s], "]" ;\n")
```

giving output like

```
"16" [shape= circle ,height= 0.5 ,width= 0.75 ,style=filled,color= grey ] ;
```

It then works out all the matings and creates small nodes for each mating and lines connecting the parents to these nodes

```
mating <- paste("\", s1, "x", s2, "\"", sep = "")  
cat(mating, "[shape=diamond,style=filled,label=\"\",height=.1,width=.1] ;\n")  
cat(paste("\", s1, "\"", sep = ""), " -> ", mating,  
  paste(" [dir=", dir, ",weight=1]", sep = ""),  
  " ;\n")  
cat(paste("\", s2, "\"", sep = ""), " -> ", mating,  
  paste(" [dir=", dir, ",weight=1]", sep = ""),  
  " ;\n")
```

# Drawing networks

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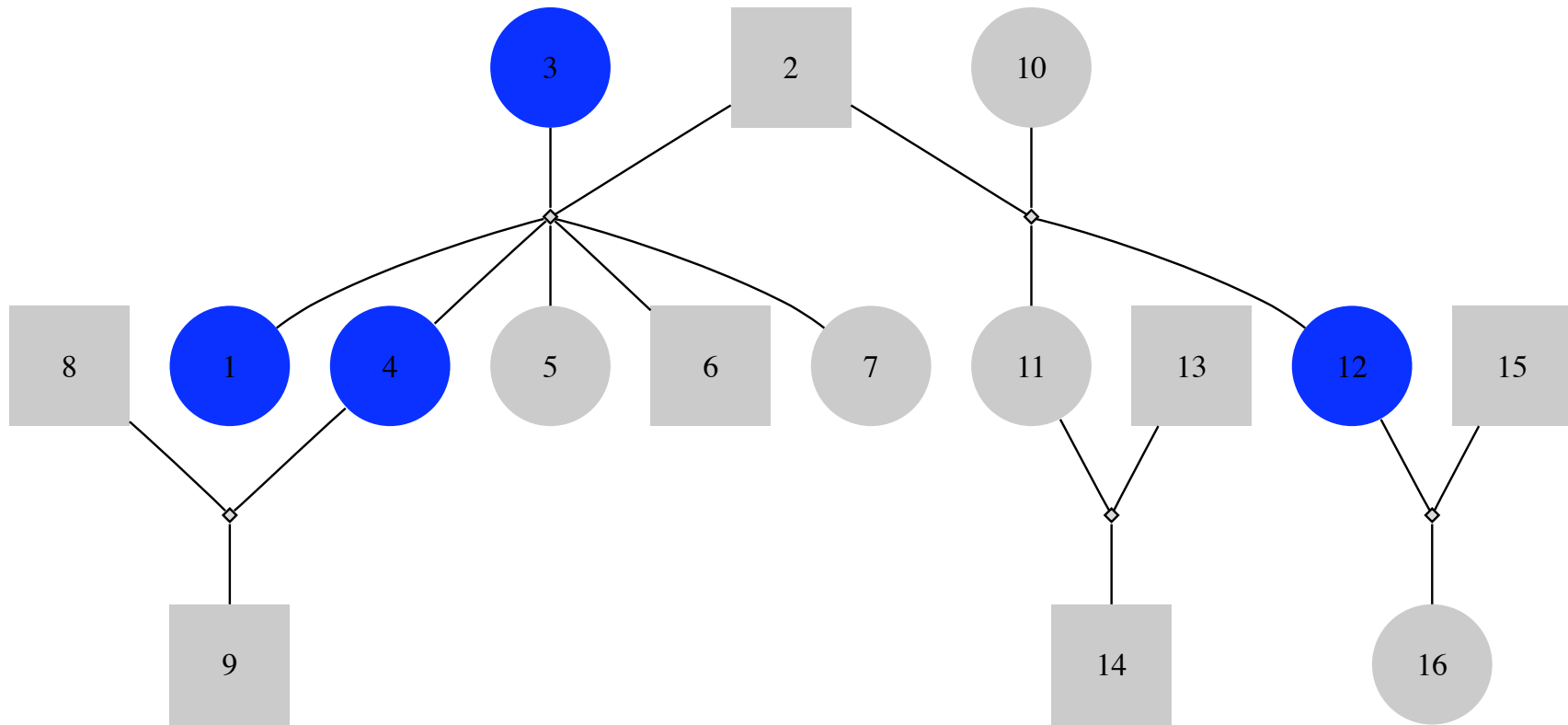
giving output like

```
"2x3" [shape=diamond,style=filled,label="",height=.1,width=.1] ;  
"2"  -> "2x3"  [dir=none,weight=1]  ;  
"3"  -> "2x3"  [dir=none,weight=1]  ;
```

and then connects children to parents.

# Drawing networks

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pedigree 10081

[Bioconductor also has GraphViz more integrated with R in the RGraphViz package]



# Chromosome simulation

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## Tasks

- Call MaCS
- Read in the lines of haploid genotypes as character strings
- Split into numbers
- Recode so 1 is the minor allele
- Combine pairs of haploids into a diploid

# Chromosome simulation

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```
makemacsdta<-function(N,length=15000,filter=0.05){
  f<-tempfile()
  system(paste("~/macs/macs",2*N,length,
    " -t .001 -r .001 2>/dev/null | ~/macs/msformatter >", f))
  input<-readLines(f)[- (1:6)]
  unlink(f)
  haplo<-do.call(rbind,lapply(strsplit(input,""),as.integer))
  diplo<-haplo[1:N,]+haplo[(N+1):(2*N),]
  af<-colMeans(diplo)/2
  diplo[,af>0.5]<- 2-diplo[,af>0.5,drop=FALSE]
  maf<-colMeans(diplo)/2
  diplo[,af<=filter,drop=FALSE]
}
```

# Chromosome simulation

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From the user's viewpoint it looks as though everything was done in R;

```
> d <- makemacsddata(1000)
> str(d)
  num [1:1000, 1:80] 0 0 0 0 0 0 0 0 0 0 0 ...
> summary(colMeans(d)/2) #maf
   Min. 1st Qu.  Median    Mean 3rd Qu.    Max.
0.00050 0.00100 0.00450 0.01142 0.01512 0.05000
```



# Chromosome simulation

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Now simulate a new version of the SKAT rare-variant test;

```
one.sim<-function(thresholds=c(Inf,1/2,1/3,1/4),  
                 sqrtweights=wuweights,  
                 N=4000, n=200, length=15000, filter=0.02)  
{  
  G <- makemacsddata(N,length,filter=filter)  
  y <- sample(rep(0:1,c(N-n,n)))  
  sapply(thresholds,  
         function(c) winskat(G,y,threshold=c,sqrtweights))  
}
```

# SVG+tooltips

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SVG (Scalable Vector Graphics) is a non-bitmap graphics format for the web.

The `RSvgDevice` and `RSVGTipsDevice` packages allow R output to SVG format.

We can use this to create graphs with links and tooltips. For example, a funnelplot showing associations between a large number of SNPs and VTE.

Point at a dot to see the SNP it represents, and click to go to information about the gene.

# SVG+tooltips

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```
for(i in 1:length(or)) {
  setSVGShapeToolTip(title=gene[i],
    desc1=snp[i],
    desc2=if(abs(lor[i]/se[i])>qnorm(0.5/n,lower.tail=FALSE))
              qvals[i] else NULL
  )

  setSVGShapeURL(paste("http://pga.gs.washington.edu/data",
                        tolower(gene[i]),
                        sep="/")
  )
  points(prec[i],lor[i], cex=1, pch=19, col='grey')
}
```

# Google Earth

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Google Earth is controlled by KML files specifying locations. KML is another plain text format.

Using `cat()`, we can write a short KML file;

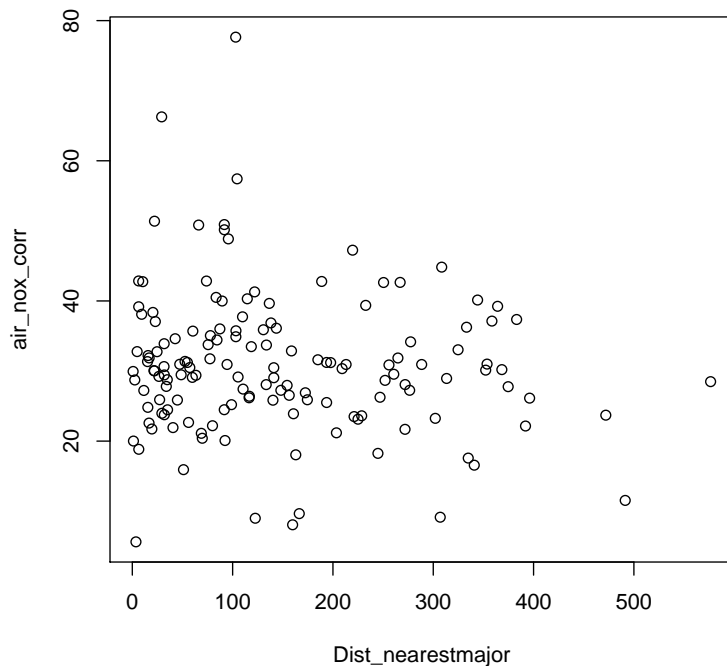
```
<?xml version="1.0" encoding="UTF-8"?>
  <kml xmlns="http://earth.google.com/kml/2.1">
    <Placemark>
      <name> 1 </name>
      <Point> <coordinates>-118.0256,34.11619,400</coordinates>
    </Point>
  </Placemark>
</kml>
```

and then send it to Google Earth with the `shell.exec(filename)` function, which opens a file using whatever is the appropriate program.

# Google Earth

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Finally, the `identify()` function lets the user select a point on a scatterplot;



In this example the points are locations where air pollution was measured, and we can call Google Earth to look at the location.