ASTROBIOLOGY Volume 19, Number 5, 2019 © Mary Ann Liebert, Inc. DOI: 10.1089/ast.2018.1835

A Maximum Subsurface Biomass on Mars from Untapped Free Energy: CO and H₂ as Potential Antibiosignatures

Steven F. Sholes, 1,2 Joshua Krissansen-Totton, 1,2 and David C. Catling 1,2

Abstract

Whether extant life exists in the martian subsurface is an open question. High concentrations of photochemically produced CO and H₂ in the otherwise oxidizing martian atmosphere represent untapped sources of biologically useful free energy. These out-of-equilibrium species diffuse into the regolith, so subsurface microbes could use them as a source of energy and carbon. Indeed, CO oxidation and methanogenesis are relatively simple and evolutionarily ancient metabolisms on Earth. Consequently, assuming CO- or H₂-consuming metabolisms would evolve on Mars, the persistence of CO and H₂ in the martian atmosphere sets limits on subsurface metabolic activity. In this study, we constrain such maximum subsurface metabolic activity on Mars using a one-dimensional photochemical model with a hypothetical global biological sink on atmospheric CO and H₂. We increase the biological sink until the modeled atmospheric composition diverges from observed abundances. We find maximum biological downward subsurface sinks of 1.5×10^8 molecules/(cm²·s) for CO and 1.9×10^8 molecules/(cm²·s¹) for H₂. These convert to a maximum metabolizing biomass of $\lesssim 10^{27}$ cells or $\le 2 \times 10^{11}$ kg, equivalent to $\le 10^{-4} - 10^{-5}$ of Earth's biomass, depending on the terrestrial estimate. Diffusion calculations suggest that this upper biomass limit applies to the top few kilometers of the martian crust in communication with the atmosphere at low to mid-latitudes. This biomass limit is more robust than previous estimates because we test multiple possible chemoautotrophic ecosystems over a broad parameter space of tunable model variables using an updated photochemical model with precise atmospheric concentrations and uncertainties from *Curiosity*. Our results of sparse or absent life in the martian subsurface also demonstrate how the atmospheric redox pairs of CO-O₂ and H₂-O₂ may constitute antibiosignatures, which may be relevant to excluding life on exoplanets. Key Words: Martian atmosphere— Photochemistry—Life on Mars—Metabolisms—Habitability—Biosignatures. Astrobiology 19, 655–668.

1. Introduction

A N OPEN QUESTION ABOUT Mars is whether it currently hosts life in its subsurface. The *Viking* landers of the 1970s have been the only direct life detection missions on Mars, and the consensus is that their results were negative (Klein, 1978, 1998). This is consistent with the poor habitability of the surface environment. Aside from plausible ephemeral briny solutions (Rennó *et al.*, 2009; Toner *et al.*, 2014; Martín-Torres *et al.*, 2015; Ojha *et al.*, 2015), the surface is cold and lacks lasting liquid water (Rummel *et al.*, 2014) to provide a solvent for life. Also, bombardment from harsh galactic cosmic ray, solar ultraviolet, and solar energetic proton radiation in the upper few meters is enough to deactivate cells over timescales of 1,000–200,000 years depending on radioresistance and regolith materials (Cockell

et al., 2000; Dartnell et al., 2007; Pavlov et al., 2012; Teodoro et al., 2018). Furthermore, highly oxidizing perchlorate in the soil can become microbicidal under martian conditions (Wadsworth and Cockell, 2017).

However, the possibility of extant martian life remains. Reported detections of transient methane (Formisano *et al.*, 2004; Krasnopolsky *et al.*, 2004; Mumma *et al.*, 2009; Webster *et al.*, 2015, 2018) have resulted in a surge of interest in the possibility of biological methanogenesis on Mars (Mickol and Kral, 2017; Yung *et al.*, 2018), although see Zahnle (2015; Zahnle *et al.*, 2011) for a dissenting view on the detections. Terrestrial microbes have also been found to use perchlorate oxidation-based metabolisms (Myers and King, 2017). Subsurface life could also use deep subsurface H₂ gas sourced through cataclastic (McMahon *et al.*, 2016), serpentinizing (Ehlmann *et al.*, 2010), or radiolytic (Onstott

¹Department of Earth and Space Sciences, University of Washington, Seattle, Washington.

²Astrobiology Program, University of Washington, Seattle, Washington.

et al., 2006) reactions as a potential energy source before reaching the surface, but there is generally no way to constrain the extent of such life based on current observables.

Instead, one way to constrain extant subsurface martian life is to assume that is in contact with the atmosphere through the porous regolith and to consider atmospheric disequilibrium. Atmospheric chemical disequilibrium is often discussed as a life-detection method for planetary atmospheres. For example, the coexistence of oxygen and methane in Earth's atmosphere is a biosignature because this out-of-equilibrium redox pair would not persist without replenishing fluxes of methane and oxygen (Lovelock, 1965; Hitchcock and Lovelock, 1967; Krissansen-Totton et al., 2016). If abiotic sources of O2 and CH4 can be ruled out, then the O₂-CH₄ pair is a disequilibrium biosignature. However, when the source of atmospheric disequilibrium is known to be abiotic, such as from photochemical or geological fluxes, and the predicted abiotic concentrations of the disequilibrium pair match that observed, then it suggests that life is not exploiting the atmospheric free energy.

Historically, it was argued that the martian atmosphere is essentially at equilibrium and therefore Mars is unlikely to support life (Lovelock, 1975, 1979, 1988). However, with modern observations, it is understood that Mars' atmosphere has the largest thermodynamic disequilibrium in the Solar System aside from Earth, with an available free energy of $\sim 136 \, \text{J/mole}$ of atmosphere (Krissansen-Totton *et al.*, 2016). This free energy is predominantly attributable to the CO-O₂ redox pair produced abiotically by the photolysis of CO₂ and H₂O in a thin, dry atmosphere.

There are compelling reasons to believe that martian life—even if it possesses radically different biochemistry to Earth life—would evolve to exploit the disequilibrium available from the redox pairs in Mars' atmosphere. The net reaction for the anaerobic CO metabolism is CO+ $H_2O \rightarrow CO_2 + H_2$ (Techtmann *et al.*, 2009) and is a relatively simple metabolism that only requires water, given a source of CO. The enzymes that catalyze this reaction on Earth, carbon monoxide dehydrogenases (CODHs), possess a variety of simple Ni-Fe or Mo active sites, suggesting that they have evolved independently multiple times (Ragsdale, 2004; Techtmann et al., 2009). In addition, the genes encoding CODH are prolific with some 6% of sequenced microbial genomes having one or more copies of the Ni-Fe CODH. This has led some to argue CO oxidation to be an evolutionarily ancient process, serving as both an energy and carbon source for the earliest forms of life in volcanic settings (Huber and Wächtershäuser, 1997; Ragsdale, 2004; Ferry and House, 2005; Lessner et al., 2006). Indeed, genomic evidence suggests the last universal common ancestor possessed genes encoding CODH (Weiss *et al.*, 2016). Also, there are modern methanogenic archaea that use CO for their entire metabolisms (Rother and Metcalf, 2004).

Aerobic CO metabolisms also exist, which oxidize CO through the net reaction $2\text{CO} + \text{O}_2 \rightarrow 2\text{CO}_2$ (Meyer and Schlegel, 1983; Ragsdale, 2004). These aerobic carboxydotrophs also use CODH enzymes that possess a variety of Mo-, Fe-, and Cu-activation sites (Jeoung and Dobbek, 2007). Given the diversity of transition metal catalysts and the abundance of CO in the martian atmosphere, it is reasonable to expect that microbial life in contact in the atmosphere would evolve to exploit this available free energy.

A much smaller portion of Mars' atmospheric thermodynamic disequilibrium, due to the small concentration of H₂, is attributable to the H₂-O₂ redox pair. Microbial methanogenesis, oxidizing H₂ with CO₂, may be one of the most primitive metabolisms on Earth (Ueno *et al.*, 2006; Weiss *et al.*, 2016; Wolfe and Fournier, 2018). Thus H₂-oxidation could also be a source of free energy for martian life, although it should be noted that CO provides >98% of the available atmospheric free energy.

All life as we know it, without exception, uses redox chemical reactions to generate energy (e.g., Falkowski et al., 2008). Given abundant free energy in the martian atmosphere, microbial life—if it exists—ought to take advantage of this "free lunch" (Zahnle et al., 2011). The thermodynamic drive to equilibrium by chemosynthetic life suggests that certain molecules in abundance, such as the CO-O₂ and H₂-CO₂ redox pairs, constitute antibiosignatures (Wang et al., 2016; Catling et al., 2018)—evidence that actively metabolizing life is absent or sufficiently meager that it does little to perturb the atmosphere beyond its abiotic photochemical state. The persistence of this photochemically induced disequilibrium and the reasonable assumption that martian life would evolve a simple CO- or H₂-metabolism to take advantage of such a disequilibrium suggests that any microbial life in communication with the atmosphere has not exploited this free lunch, and thus it represents an antibiosignature.

Disequilibrium in Mars' atmosphere has previously been used to constrain a maximum extant biomass in the subsurface using a modified photochemical model to include biogenic sinks on CO and H₂ (Weiss *et al.*, 2000). However, this study was potentially inaccurate because it used fixed values for photochemical model parameters tuned to modern abiotic martian conditions, whereas one should use values that account for their large uncertainties. Instead, given the uncertainties on these parameters (described in Section 2.2), they could be tuned differently, but within allowable error bars, to an assumed Mars with biogenic sinks on CO or H₂. This procedure could potentially alter the outcome for calculated subsurface biomass.

In this study, we consider how much biomass could exist in the current subsurface and be feeding off atmospheric redox couples. A subsurface environment is assumed because persistent liquid water on Mars today can only exist in the subsurface where the geothermal gradient allows an aquifer (*e.g.*, Mellon *et al.*, 1997; Feldman *et al.*, 2004; Grimm and Painter, 2009; Dundas *et al.*, 2014).

We use a novel approach by optimizing the photochemical model over a broad range of tunable parameter space that assumes a biosphere for calculating unknown variables. In addition to the significance of the optimization procedure, we additionally improve and expand upon the work done by Weiss *et al.* (2000) in four major ways: (1) we include sources and sinks for all metabolic reactants and products as opposed to only sinks on CO and H₂. (2) We use updated, more precise present day atmospheric compositions collected by the *Curiosity* rover with an improved photochemical model. (3) We account for multiple combinations of the plausible chemoautotrophic ecosystems that use different net metabolic pathways and use improved estimates for cellular maintenance energy. (4) Because the delivery of substrate gases to subsurface life (and hence its biomass)

might be restricted by downward diffusion through the regolith, we develop an improved gas diffusion model and test over a broad range of surface properties and crustal gradients. Thus, our comprehensive results provide rigorous constraints on the maximum biomass (in communication with the atmosphere) that could exist on Mars today.

2. Methods

To investigate the effects of biological sinks on the martian atmosphere, we use a one-dimensional (1D) photochemical code originally developed for modeling the early Earth by Kasting (1979), but our nominal model has since been modified and validated for the modern martian atmosphere (Zahnle et al., 2008; Smith et al., 2014; Sholes et al., 2017). This model is built to include C-H-O-N-S (carbon, hydrogen, oxygen, nitrogen, and sulfur) chemistry and improves upon the Nair et al. (1994) model used by Weiss et al. (2000) in three major ways: (1) for redox conservation, hydrogen escape is balanced by an abiotic tunable deposition velocity (v_{dep}) of reactive oxidizing species to the surface rather than arbitrary O escape at the top of the model (Lammer et al., 2003; Zahnle et al., 2008; Lillis et al., 2017); (2) the implementation of diffusion-limited hydrogen escape, both of which are justified by Zahnle et al. (2008) with reference to data and models and by measurements by the Mars Atmosphere and Volatile EvolutioN (MAVEN) mission (Jakosky et al., 2018) where the measured H escape rate of $0.8-7.6\times10^8$ atoms/(cm²·s) encompasses the estimated diffusion-limited escape rate of $(3.3\pm1.1)\times10^8$ H atoms/(cm²·s) within uncertainties (Catling and Kasting, 2017, p. 148); and (3) implementing a lower boundary condition where the CO₂ concentration is no longer fixed.

With the exception of the CO₂ treatment (described in more detail below), we follow the same parameters as the aforementioned models. Atmospheric surface pressure is set to 6.5 mbar, close to present day global average levels (Haberle et al., 2008). The model uses uniformly spaced 1 km resolution grids up to 110 km. Ionospheric chemistry is not directly modeled, but downward fluxes at the top of the model of key photolytically produced ionospheric species are included, namely NO, N, and CO. Oxygen escapes out the top of the model at a rate of 10^7 O atoms/(cm²·s) following (Zahnle et al., 2008) and is of the same order as measurements made by MAVEN [$\sim 3 \times 10^7$ O atoms/(cm²·s)] (Jakosky et al., 2018). Vertical transport of species is dominated by eddy diffusion. The temperature profile is approximated by $T = T_s - 1.4 h$ for the lower 50 km and isothermal above; T_s is the surface temperature (detailed below) and his the height above the surface (in km). Water vapor is kept at a constant relative humidity in the lower atmosphere to produce the observed 9.5 pr-µm (Zahnle et al., 2008).

Previous versions of the photochemical model held CO₂ at a constant mixing ratio, assuming it to be replenished by indefinitely large surface (e.g., polar caps) and subsurface reservoirs (e.g., Zahnle et al., 2008). In this study, we allow the CO₂ mixing ratio to vary, rather than be fixed at a set concentration. This allows for more accurate behavior when biology is included because some metabolisms release or draw down CO₂. In particular, CO and O₂ are replenished through the photolysis of CO₂, so even though they would be drawn down as sinks in different metabolisms, they

would be artificially replenished by an unrealistic injection of CO_2 (up to the set value) at all grids and time steps if CO_2 were fixed numerically in the model.

Validation tests were successfully performed to ensure that the updated model, where the CO₂ concentrations can vary, matches the previously validated models in their atmospheric structure and composition. Additional tests were done to compare each modeled metabolism with and without a fixed CO₂ mixing ratio (for both the "fixed parameter" and "optimized parameter" models described below). No significant differences were found other than a decrease in the maximum allowable sink for the net aerobic carboxydotrophy metabolism. When CO₂ concentrations are fixed, the maximum allowable biogenic sink for the net aerobic carboxydotrophy metabolism is unrealistic as it approaches the column-integrated production limit of CO due to the artificial replenishment of CO and O₂ in the atmosphere. All work presented in this study reflects the improved model with CO₂ concentrations no longer fixed, thus more accurately modeling the effect of each metabolism.

The influence of possible subsurface biological metabolic activity is modeled by ground-level fluxes in the model. For each of the metabolic ecosystems simulated, the model is run with that metabolism's substrate gases removed directly from the atmosphere by a fixed downward surface flux, while we simultaneously inject its metabolic products directly into the atmosphere at the lower boundary. The model is then run to steady state and ground-level mixing ratios are compared with the observed abundances measured for the modern martian atmosphere (Table 1) (Krasnopolsky and Feldman, 2001; Webster et al., 2015, 2018; Franz et al., 2017). This process is repeated as the surface fluxes are incrementally increased until the modeled atmosphere diverges from the current atmospheric compositions within a 2σ (95%) uncertainty. We assume that this divergence then sets the limit on the extent of a metabolizing subsurface biomass because its activity would not violate the known atmospheric composition.

2.1. Fixed parameter model

We use the aforementioned nominal Mars model, which has been tuned and validated against the modern martian atmosphere (Zahnle *et al.*, 2008; Smith *et al.*, 2014; Sholes *et al.*, 2017). There are five possible net chemoautotrophic metabolisms that could feasibly be living off atmospheric energy. We express these metabolisms as both individual metabolic pathways and combined metabolic ecosystems as the Gibbs free energy produced through either net reaction is identical. The metabolisms (Mets.) are as follows:

• Met. 1: Anaerobic CO metabolism only:

$$CO + H_2O \rightarrow CO_2 + H_2 \tag{1}$$

• Met. 2: Methanogenesis only:

$$CO_2 + 4H_2 \rightarrow CH_4 + 2H_2O$$
 (2)

• Met. 3: Anaerobic CO metabolism and methanogenesis (*i.e.*, 4×Met.1 + Met. 2):

$$4CO + 2H_2O \rightarrow 3CO_2 + CH_4 \tag{3}$$

Species	Observed mixing ratio	Uncertainty (1σ)	References	
$\overline{\mathrm{O}_2}$	1.74×10^{-3}	6×10^{-5}	Franz <i>et al.</i> (2017)	
${ m CO}_2$	7.47×10^{-4}	2.6×10^{-6}	Franz <i>et al.</i> (2017)	
	1.5×10^{-5}	5×10^{-6}	Krasnopolsky and Feldman (2001)	
H ₂ CH ₄	6.9×10^{-10}	1.3×10^{-10}	Webster et al. (2015) ^a	
CO_2	0.96	7×10^{-3}	Mahaffy et al. (2013)	

TABLE 1. MARS' ATMOSPHERE: MODERN MARTIAN ATMOSPHERIC COMPOSITION AND UNCERTAINTIES FOR MODELED FLUXES

Here we use the slightly higher background value of Webster et al. (2015) rather than that from Webster et al. (2018) for a more conservative upper limit on biomass.

^aThese measurements are of a mean atmospheric level of 0.69 ppb and are controversial (see Zahnle *et al.*, 2011), thus we place an upper bound of 9.4×10^{-10} representing a 2σ detection.

• Met. 4: Aerobic carboxydotrophy or an equivalent net anaerobic CO metabolism, methanogenesis, and methanotrophy ecosystem [i.e., $4 \times \text{Met.}1 + 2 \times \text{Met.}2 + \text{methanotrophy}$ (CH₄ + 2O₂ \rightarrow 2H₂O + CO₂)]:

$$2CO + O_2 \rightarrow 2CO_2 \tag{4}$$

• Met. 5: Aerobic hydrogenotrophy or an equivalent net methanogenesis and methanotrophy ecosystem (*i.e.*, Met.2 + methanotrophy):

$$O_2 + 2H_2 \rightarrow 2H_2O \tag{5}$$

These end-member cases represent the full range of possible martian ecosystems subsisting off atmospheric free energy. Weiss et al. (2000) only modeled Met. 4 and Met. 5 as they believed the most energetic reactions would likely use O₂ as an oxidant. We do not consider a methanotrophyonly metabolic system (or combinations of ecosystems that include methanotrophs without methanogens) because it is infeasible given that the concentration of background methane is <1 ppb (Webster et al., 2015, 2018) and thus will produce a much smaller maximum biomass than other metabolisms. Reported transients of methane >10 ppb are localized and controversial in nature (Zahnle et al., 2011). Notably, in the *Curiosity* tunable diode laser results of Webster et al. (2018), such transients are only found sometimes in a direct measurement protocol, but never found in a more sensitive enrichment protocol. Thus, these results demand skepticism as the transients appear to be more of a function of measurement protocol than of Mars. Similarly, we do not test metabolisms that produce gases not observed in the current martian atmosphere (e.g., sulfate reducers producing H_2S).

2.2. Optimized parameter model

The methodology for what we call the "fixed parameter" model (Section 2.1) may not provide a rigorous maximum biogenic sink because it is tuned to a modern abiotic Mars with fixed unknown parameters (described below) that have been tuned in the model. If life is actively metabolizing on Mars, then the tuned parameter values could be different to what is commonly assumed, and their incorrect tuning could mask biogenic sources/sinks. Thus, the "fixed parameter" model underestimates the maximum biogenic sink.

We have identified three main tunable variables in the nominal model where the value has significant impact on the atmospheric composition and is either unknown or not agreed upon in the literature: surface temperature, ionospheric flux, and deposition velocity. Of these, mean surface temperature is the most constrained based on observed measurements, but global mean surface temperature is dependent on factors such as global circulation model integrations and atmospheric dust content (Haberle, 2013). Ionospheric fluxes of the odd nitrogen species (NO and N) are dependent on their concentrations and ratio, which are not entirely known. Krasnopolsky (1993) considers cases where the upper boundary flux of NO into the neutral atmosphere is both nonexistent and appreciable [10' molecules/(cm 2 ·s)], so we consider the range 10^1 – 10^8 molecules/ $(cm^2 \cdot s)$ consistent with sensitivity testing by Smith et al. (2014). A flux of CO into the upper atmosphere is set to be equal to that of NO to conserve redox balance, while N is set to 10% that of NO (Zahnle et al., 2008; Sholes et al., 2017).

Deposition velocity of reactant species is the dominant free parameter of the model. It is a constant used to simulate mixing, molecular diffusion, adsorption, and reactions of species with the surface regolith in a single variable. Higher v_{dep} indicates a more chemically reactive species (Seinfeld and Pandis, 2016; Catling and Kasting, 2017). In a physical system, the deposition velocity for each species would be different, but these variations would be small, so a single v_{dep} is assumed for all reactive species. This is considered a good approximation (Zahnle et al., 2008) as H₂O₂ and O₃ are the primary reactants that must have the v_{dep} tuned for each model run to produce the modern abundances for the primary atmospheric constituents and computational restrictions prevent optimization of v_{dep} over 30+ species. CO₂, CO, O₂, and H₂ are considered nonreactive with the surface and their $v_{dep} = 0$ cm/s (See Zahnle et al., 2008 for more on O2 dry deposition and Sholes et al., 2017 for a review on CO deposition). For each species, v_{dep} is a fixed value, but is used to compute a variable surface flux (Φ) for each chemically reactive species, which depends on species abundance as $\Phi = -v_{dep} n_{i,surf}$, where $n_{i,surf}$ is the surface number density for species i (Seinfeld and Pandis, 2016; Catling and Kasting, 2017). This net oxidant sink balances the atmospheric redox budget by countering hydrogen escape to space, which oxidizes the atmosphere (Lammer et al., 2003; Zahnle et al., 2008; Seinfeld and Pandis, 2016). The biological consumptions of CO, O₂, and H₂ are included using a separate fixed flux term.

To test how optimizing the tunable parameters affects the maximum biological sink in the modern martian atmosphere, we perform a grid search and optimization procedure through the parameter space for the five net ecosystems described in Section 2.1. We incrementally increase the net metabolic source/sink fluxes as described for the nominal model, but at each increment, we perform a grid search of model parameter space to find whether any possible set of parameters (for temperature, ionospheric flux, and deposition velocity) can reproduce the observed martian atmosphere, while incorporating the biogenic fluxes, what we describe as our "optimized parameter" model. At every grid point in parameter space, we perform an optimization procedure (scipy.optimize.minimize in Python) to ensure that all regions in parameter space were explored and not missed by the coarse grid search. Table 2 summarizes the plausible parameter ranges, bounds, and number of grid points for each parameter, in addition to the tuned values that produce the modern atmospheric concentrations.

The optimization procedure minimized the Chi-Squared (χ^2) value comparing the modeled atmospheric composition with the measured atmospheric composition and uncertainties in Table 1. This procedure was continued until the modeled atmospheric compositions diverged from the observed composition, regardless of parameter values.

2.3. Atmospheric diffusion

Once maximum permissible biogenic sinks are found in both the "fixed parameter" and "optimized parameter" versions of the model, we consider how these fluxes compare with passive atmospheric diffusion into the crust. This is because the dry, harshly irradiated, and constantly oxidized martian surface is ostensibly uninhabitable (see Section 1), so any extant life would need to take advantage of aquifers in the subsurface. There may be small pockets of briny fluids in the upper few kilometers (Orosei et al., 2018), but a deep watersaturated layer should exist within 10 km of the surface as a consequence of the geothermal gradient (Clifford, 1993; Clifford et al., 2010; Michalski et al., 2013). In this study, we calculate to what depth microbes could plausibly exploit the atmospheric free energy and thereby assign a maximum crustal depth to our upper biomass limit (i.e., the biomass of microbes living below this depth would be limited by diffusion of substrate gases and would therefore have smaller surface sinks than the calculated maximum fluxes that ignore a diffusion restriction).

To calculate the diffusion flux of CO and H_2 at depth (the potential flux), we use a modified form of Fick's laws, which assume mean free paths of diffusing gases are greater than the typical pore size such that Knudsen diffusion dominates. In one dimension, the general form of Fick's second law states the following:

$$\frac{\partial n_i}{\partial t} = \frac{\partial}{\partial z} \left(D_i(z) \frac{\partial n_i}{\partial z} \right) \tag{6}$$

where n_i is the number density for species i (molecules/ cm³), z is the depth (cm), t is time (s), and D_i is the diffusion coefficient (diffusivity, cm²/s). In steady state, the concentrations will not change with time, so the left-hand side of this equation is set to 0. The solution for the concentration gradient, $\frac{\partial n_i}{\partial z}$, can then be used to find the potential flux, F_i , by Fick's first law:

$$F_i = -D_i(z) \frac{\partial n_i}{\partial z} \tag{7}$$

The physical environment over which the gas diffuses, changes greatly through the upper $\sim 10 \, \mathrm{km}$ of martian regolith. This means that the diffusivity, the controlling factor, changes drastically with increasing depth due to increasing pressure and temperature, and can be written as follows:

$$D_i(z) = \frac{\varepsilon(z) \, r(z)}{3 \, \tau(z)} \sqrt{\frac{8 \, R \, T(z)}{\pi \, m_i}} \tag{8}$$

where $\varepsilon(z)$, r(z), $\tau(z)$, and T(z) are the porosity, pore size (cm), tortuosity (path twistiness), and temperature (K) at a given depth. R is the gas constant and m_i is the molar mass of the species. We follow Stevens et al. (2015, 2017) in assuming that porosity and tortuosity decrease exponentially with depth with $\varepsilon(z) = \varepsilon_0 e^{\frac{-z}{k}}$ and $\tau(z) = \tau_0 e^{\frac{-z}{3k}}$, where ε_0 and τ_0 are the surface porosity and tortuosity, respectively, and k is a scaling factor set by the pore closure depth (Clifford, 1993). Temperature is modeled as a linear geothermal gradient, $T(z) = a_T z + T_0$, where T_0 is the modeled surface temperature and a_T is the gradient (Michalski et al., 2013). We assume pore size follows a constant pressure gradient (Stevens et al., 2015), $r(z) = a_r z + r_0$, where a_r is the gradient defined as $\frac{r_0}{-k}$ such that pore size is 0 at the pore closure depth and r_0 is the average surface pore size. We perform these calculations over a range of plausible gradients and surface parameter values (detailed below).

If we consider a layer of biota sustaining off atmospheric energy, living at a depth z_b , and metabolizing all available reactants (CO or H₂), Equation 6 can be numerically solved as a boundary value problem. Using observed surface number densities, $n_i(0)$, and setting $n_i(z_b) = 0$; we numerically solve for n(z) (scipy.integrate.solve_bvp in Python) in steady state. We can then use Equation 7 to solve for potential flux as a function of biotic layer depth (z_b) . In practice, microbes cannot metabolize with substrate conditions of zero within the

TABLE 2. PARAMETER SPACE: UNKNOWN TUNABLE MODEL PARAMETERS AND THEIR ASSUMED RANGES FOR MARS

Parameter	Plausible range	Grid space	Tuned valued	References
Mean surface temperature, T_s (K) Surface deposition velocity, V_{dep} (cm/s) Ionospheric flux, $\varphi_{NO,CO}$ [molecules/(cm ² ·s)]	$199-215 \\ 0.001-0.1 \\ 10^{1}-10^{8}$	5 3 5	$ \begin{array}{c} 210 \\ 0.012 \\ 7.3 \times 10^2 \end{array} $	Haberle (2013) Zahnle <i>et al.</i> (2008) Krasnopolsky (1993); Smith <i>et al.</i> (2014)

Tuned values are for the fixed parameter abiotic modern Mars model. Grid space refers to how many equally spaced values of the parameter were used in the optimization grid search within the plausible range for the optimized parameter model. v_{dep} for CO, CO₂, H₂, and O₂ are set to 0 cm/s. The tuned v_{dep} parameter refers to that assumed for all reactive species, including both O₃ and H₂O₂.

biologically active layer, but observations of Antarctic soils have found that microbes can metabolize with CO concentrations of 20 ppb (Ji *et al.*, 2017). Setting $n_i(z_b)$ to the equivalent of 20 ppb for CO under Antarctic conditions (approximated as 1 bar and 255 K) results in nearly identical potential fluxes as the $n_i(z_b) = 0$ case demonstrating that a zero-concentration lower boundary condition is a reasonable assumption.

Given the uncertainties of the physical surface conditions and gradients, we employ a Monte Carlo simulation to test the potential flux under both a range of z_b and physical parameter values (for porosity, tortuosity, average pore size, pore closure depth, and temperature). For each possible z_b , spaced every 10 m between 0 and 10 km, we perform 10,000 diffusion calculations, where we assume each parameter has a uniform distribution bounded by the parameter ranges described above and sample these distributions randomly. From these outputs, we obtain a likelihood distribution for the diffusion flux as a function of biological layer depth.

Surface porosity ranges from ε_0 =0.2–0.6, with low values characteristic of lunar estimates and high values approximating *Viking* lander observations (Clark *et al.*, 1976; Clifford, 1993; Sizemore and Mellon, 2008). Estimated surface tortuosity ranges from τ_0 =1.5–2.5 as measured in Marsregolith simulant soil (Sizemore and Mellon, 2008). We take an average surface pore size range of r_0 =10⁻³–10⁻⁴ cm based around the typically cited value of 6×10^{-4} cm (Weiss *et al.*, 2000) and experimental pore size distributions using glass beads (Sizemore and Mellon, 2008). The pore closure depth due to compaction ranges from k=6–26 km depending on how saturated the crust is with water. The extremes are most likely overestimates/underestimates and a value of 10 km is typically assumed (Hanna and Phillips, 2005). We test temperature gradients of 10–30 K/km (Michalski *et al.*, 2013).

3. Results

Figure 1 shows the effects of increasing the subsurface biological sinks for Mets. 1–3, while Fig. 2 shows the results for Mets. 4 and 5. In all cases, as the sink is ramped up, the resulting modeled atmospheric composition eventually diverges from the 2σ uncertainty of the measured modern atmospheric composition. For example, in the fixed parameter model Met. 1 case, the CO abundance falls below the observed 2σ abundance of 744 ppm at a downward biologic flux of 8.4×10^5 CO molecules/(cm²·s). In this case, CO is considered the "break species" as CO diverges from the observed concentrations at a lower biological sink than the other measured constituents (O₂, H₂, CH₄, and CO₂). The flux at which this divergence occurs is the maximum permissible flux where the modeled metabolism can still replicate observed abundances and is listed in Table 3 for the fixed parameter model Met. 1 case.

In this example (Met. 1, anaerobic CO metabolism, fixed parameter model), as the biological sink on CO is ramped up, CO concentrations decline, O_2 abundances steadily rise, and CO_2 abundances eventually decline as well. These behaviors are the result of the simulated biological drawdown on CO. CO_2 is readily converted into CO and O_2 through photolysis, thus removal of CO through high biological sinks inhibits the recombination reactions back into CO_2 and the injection of CO_2 as a product of the metabolism eventually becomes in-

sufficient to counteract the removal of CO. This leads to low CO, excess O₂, and eventually low concentrations of CO₂ (Catling and Kasting, 2017 pp. 338–340).

Except for metabolisms that produce CH_4 , in all other metabolisms in the fixed parameter model runs, CO concentrations deviate from observations at the lowest flux levels (CH_4 deviates at lower fluxes in metabolisms that produce it). This is due to either the direct biogenic sink on CO or the removal of O_2 (a byproduct of CO_2 photodissociation), which throttles the recombination of CO_2 and leads to excess CO. Because CO has the smallest observational uncertainty, the nature of the χ^2 -calculation in the optimized parameter model will end up attempting to constrain CO more precisely than the other species. Thus, the optimized parameter model runs generally have CO_2 concentrations that deviate at lower fluxes, except for CH_4 -producing metabolisms and the aerobic H_2 -metabolism (where O_2 and H_2 are drawn down and eventually diverge from observations).

The maximum biogenic sink fluxes for each of the net ecosystems with both the fixed and optimized parameter models are summarized in Table 3. We find that the maximum downward flux on CO or H_2 ranges from 10^5 to 10^8 molecules/(cm²·s). As expected, the optimized version of the model produced maximum fluxes that were consistently higher than their fixed parameter model counterparts. The exceptions are Mets. 2 and 3, which had comparable maxima due to CH_4 building up too high and the tunable parameters unable to effectively remove CH_4 . The anaerobic H_2 -metabolism (Met. 5) allowed for the greatest downward biogenic sink of 1.9×10^8 H_2 molecules/(cm²·s) in the optimized parameter model with the aerobic CO-metabolism (Met. 4) having a similar maximum of 1.5×10^8 CO molecules/(cm²·s).

Our fixed parameter model results show a maximum downward sink of 2×10^6 CO molecules/(cm²·s) for Met. 4 and 4×10^7 H₂ molecules/(cm²·s) for Met. 5, which are, respectively, 30 times smaller and 2 times greater than the maximum sinks from Weiss *et al.* (2000) (which tested only these 2 metabolisms). While it is expected, given our model improvements, that we find an improved result for Met. 4, it may seem counterintuitive that we find a slightly greater sink for Met. 5 (methanogenesis and methanotrophs) compared with Weiss *et al.* (2000). This stems from our use of 2σ uncertainties on atmospheric abundances rather than an arbitrary 25% of the observations (the ratio of the measured uncertainty to the value) they used [at 1σ , we find a maximum sink of $\sim 2 \times 10^6$ H₂ molecules/(cm²·s) for the fixed model Met. 5].

Maximum biogenic fluxes can be used to estimate maximum extant biomass. The conversion from flux to biomass is done using the following equation:

$$M_{\text{max}} = \frac{\Phi_{\text{max}} A_{Mars} \Delta G}{q_{BPR} N_A} \tag{9}$$

Here, the maximum metabolizing biomass $(M_{\rm max})$ is equal to the maximum biogenic sink in model unit $[\Phi_{\rm max};$ molecules/ $({\rm cm}^2 \cdot {\rm s})]$ times the surface area of Mars $(A_{Mars};$ cm²) over the ratio of the basal power requirement (BPR) for life $[q_{BPR};$ kJ/ $({\rm s\cdot cell})]$ to the specified reaction free energy $(\Delta G;$ kJ/mol) for each metabolism times Avogadro's number $(N_A;$ molecules/mol). The Gibbs free energies for each equation are $-24\,{\rm kJ/mol}$ CO, $-24\,{\rm kJ/mol}$ H₂, $-48\,{\rm kJ/mol}$ CO, $-264\,{\rm kJ/mol}$

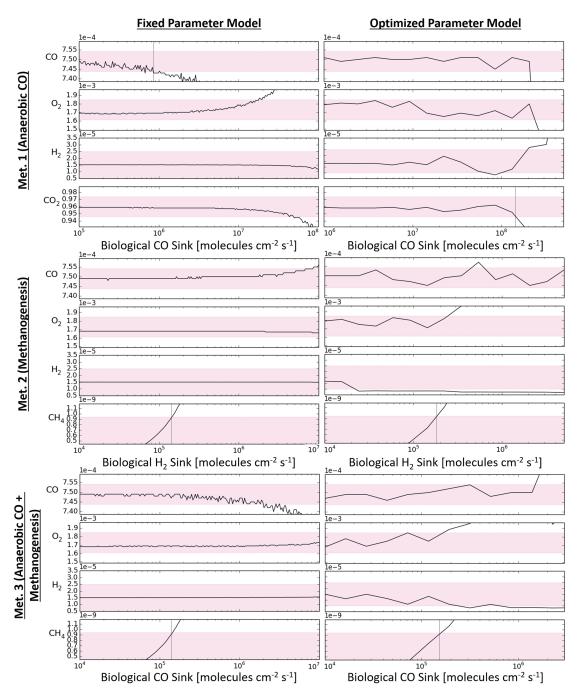


FIG. 1. Modeled atmospheres for Mets. 1–3 as biogenic sinks are incrementally ramped up. Vertical axes are gas mixing ratios. Left panels are for the fixed parameter model and the right panels for the optimized parameter version (see Secs. 2.1 and 2.2). Shaded regions represent a 2σ uncertainty for the mixing ratios of CO, O₂, H₂, CH₄, and CO₂. Vertical dotted lines indicate where the model diverges from observations (max biological sink). CO₂ concentrations do not vary for Met. 2 within or below this biogenic sink range. Met., metabolism.

mol CO, and $-215 \,\text{kJ/mol}$ H₂ for Equations 1–5, respectively (calculated with the freely available database model of Krissansen-Totton *et al.*, 2016, and using the nominal surface conditions and gas concentrations for Mars).

Quantifying the minimal BPR of organisms is currently the subject of much debate because all the contributing factors are not well characterized (see Section 4) (Hoehler and Jorgensen, 2013; LaRowe and Amend, 2015; Lever *et al.*, 2015; Kempes *et al.*, 2017; Bradley *et al.*, 2018). Many estimates

have been made, but we elect to use the value of 3×10^{-23} kJ/(s·cell) from Lever *et al.* (2015) and LaRowe and Amend (2015). This value is the smallest measured BPR from measurements of sulfate-reducing bacteria in anoxic marine sediments (Lever *et al.*, 2015). However, the BPR could potentially be as low as 1×10^{-24} kJ/(s·cell), which is a theoretical limit to prevent racemization of amino acids. BPRs less than this value may be able to sustain individual cells, but are characterized by population decay and thus not

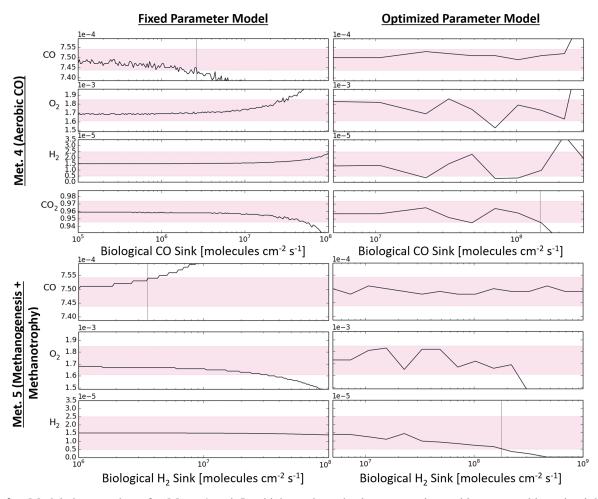


FIG. 2. Modeled atmosphere for Mets. 4 and 5, which produce the largest maximum biomass, as biogenic sinks are incrementally ramped up. Vertical axes are gas mixing ratios. Left panels are for the fixed parameter model and the right panels for the optimized parameter version (see Secs. 2.1 and 2.2). Shaded regions represent a 2σ uncertainty for the mixing ratios of CO, O₂, H₂, CH₄, and CO₂. Vertical dotted lines indicate where the model diverges from observations (max biological sink). CO₂ concentrations do not vary for Met. 5 within or below this biogenic sink range.

appropriate for characterizing a robust minimum biomass (one that is neither growing nor decaying).

Using the minimum BPR value of Lever et al. (2015), we find an upper limit of $\sim 10^{27}$ cells that could be supported of the available free energy of the martian atmosphere. This corresponds to the maximum biogenic sink for both the optimized parameter Met. 4 (aerobic carboxydotrophs) and Met. 5 (aerobic hydrogenotrophs) models. These provide similar maximum biomass values as their slightly varying max fluxes and free energy converge on a similar maximum biomass based on Equation 6. Additional conversions from this estimated cellular biomass into a metric biomass are provided in Section 4.

The likelihood functions for our Monte Carlo simulation on diffusion of CO and H_2 into the subsurface are plotted in Fig. 3. The depths at which these potential fluxes equate to our calculated maximum downward biogenic sinks $[2.1\times10^8$ molecules/(cm²·s) for CO-sink metabolisms and 1.3×10^8 molecules/(cm²·s) for H_2] represent the maximum plausible depth of any microbe subsisting off atmospheric free energy. The intercept depths for the CO-sink and H_2 -sink metabolisms with the median flux distribution are ~ 6.5 and ~ 1 km, respectively. Below these depths, the limiting factor for de-

termining the maximum extant biomass is the flux of reactants rather than the photochemistry.

4. Discussion

Our calculated maximum extant metabolizing martian biomass of $\sim 10^{27}$ cells is difficult to interpret in isolation and so we compare it to Earth's total biomass. Typical estimates for Earth's biomass are classically given in terms of petagrams of carbon mass (Pg C= 10^{15} g C) and range from 550 Pg C (Bar-On *et al.*, 2018) to 720 Pg C (Kallmeyer *et al.*, 2012). Converting these values into an estimated number of cells requires an assumption on the average cellular carbon weight. Average cellular carbon mass ranges from 5 to 85 fg C/cell¹ for microbial cells (Bakken, 1985; Kallmeyer *et al.*, 2012), so an estimated total number of cells on Earth ranges from $\sim 6.5 \times 10^{30}$ to $\sim 1.4 \times 10^{32}$ cells. Thus, our calculated maximum cellular biomass estimate of 10^{27} cells is $2-40 \times 10^{-5}$ of Earth's biomass or roughly one hundred thousandth of the estimated total of Earth's biomass.

Converting this maximum cellular biomass for Mars into a metric biomass (in kg) requires an average microbial mass.

Results: Maximum Allowable Downward Flux for Each Metabolism and the Equivalent Biomass ω. TABLE

	$Biomass \ (kg)$	$ 1.8 \times 10^{10} \\ 2.5 \times 10^{7} \\ 4.2 \times 10^{7} \\ 2.2 \times 10^{11} \\ 2.2 \times 10^{11} $
Optimized	Biomass (cells)	$ \begin{array}{c} 2.6 \times 10^{26} \\ 3.5 \times 10^{23} \\ 5.9 \times 10^{23} \\ 3.1 \times 10^{27} \\ 3.2 \times 10^{27} \end{array} $
	Break Species	CO ₂ ↓ CH ₄ ↑ CH ₄ ↑ CO2 ↓ H2 ↓
	Max flux $[molecules/(cm^2 \cdot s)]$	1.4×10 ⁸ 1.9×10 ⁵ 1.6×10 ⁵ 1.5×10 ⁸ 1.9×10 ⁸
Fixed	Biomass (kg)	$ \begin{array}{c} 1.1 \times 10^{8} \\ 1.8 \times 10^{7} \\ 3.7 \times 10^{9} \\ 5.1 \times 10^{9} \end{array} $
	Biomass (cells)	$ 1.6 \times 10^{24} \\ 2.6 \times 10^{23} \\ 5.2 \times 10^{23} \\ 5.3 \times 10^{25} \\ 7.3 \times 10^{25} $
	Break Species	CO↓ CH4↑ CH4↑ CO↓
	Max flux $[molecules/(cm^2 \cdot s)]$	8.4×10 ⁵ 1.4×10 ⁵ 1.4×10 ⁵ 2.6×10 ⁶ 4.4×10 ⁶
	Net reaction	$CO + H_2O \rightarrow CO_2 + H_2$ $CO_2 + 4H_2 \rightarrow CH_4 + 2H_2O$ $4CO + 2H_2O \rightarrow 3CO_2 + CH_4$ $2CO + O_2 \rightarrow 2CO_2$ $O_2 + 2H_2 \rightarrow 2H_2O$
	Metabolism	Anaerobic CO-only Methanogenesis Anaerobic CO and methanogenesis Aerobic CO-only Methanogenesis and methanotrophy
	No.	10 K 4 K

Biomass estimates are based on basal power requirements of 3×10^{-23} kJ/(s·cell) (LaRowe and Amend, 2015; Lever et al., 2015) and an average cellular mass of 7×10^{-17} kg (Bratbak and Dundas, 1984; Hoehler and Jorgensen, 2013). The columns headed "Break Species" describe which atmospheric species deviate from observed abundance first and whether they are higher or lower than the 2 σ detections. The greatest biomass estimate model runs are highlighted in bold.

Assuming an average dry mass of 14 fg/cell (Kallmeyer et al., 2012; Hoehler and Jorgensen, 2013), this amounts to $\sim 10^{11}$ kg total mass, assuming that dry cell mass constitutes 20% of total cellular mass (Bratbak and Dundas, 1984). In more tangible terms, this total mass is equivalent to ~ 1 million blue whales (taking the average mass of a blue whale as 1.85×10^5 kg). While this amount of blue whales worth of microbial life on Mars appears large, it is vanishingly small compared to the total biomass on Earth today and is an upper limit based on generous assumptions that give the largest biomass.

As a comparison to the previous estimate, Weiss *et al.* (2000)'s work produced maximum biomass of $\sim 10^{10}$ kg or $\sim 120,000$ blue whale masses. However, this value uses their calculated energy flux at 10 m depth, assumes only 10% metabolic efficiency, and uses a "typical maintenance energy" value of $\sim 10^{-4}$ kJ/(g·s) [$\sim 10^{-18}$ kJ/(cell·s) using our assumed average cell mass]. While it may be reasonable to assume such a power requirement and biological efficiency, this does not place an upper limit on the maximum possible biomass as microbes have been discovered subsisting off smaller energy fluxes. Indeed, if we take the maximum flux values used in their biomass calculations and apply our updated values for BPR, full metabolic efficiency, and updated Gibbs free energy, then their maximum biomass would be $\sim 10^{29}$ cells equivalent to $\sim 10^{13}$ kg or ~ 70 million blue whale masses.

The maximum biomass we calculate is likely an overestimate. There are other considerations that we have not included in this methodology that would lower the upper bound on biomass. It is assumed here that the metabolisms work at 100% efficiency—which is unlikely as energy is wasted in "spillover" reactions and Darwinian evolution favors survival over efficiency. We also use BPRs to estimate biomass, but this is the minimum energy required to keep cells alive and does not account for additional energy costs such as reproduction. Abiological oxidation reactions with the regolith should contribute a small portion of our calculated maximum reactant sinks, thus the maximum biological sinks would be smaller than reported in this study.

There is also a great uncertainty in the minimum power requirements necessary to sustain actively metabolizing life. Older literature typically used the term "minimum maintenance energy" (including Weiss *et al.*, 2000), which is defined as the minimum energy flux required to sustain a steady-state population without growth. However, recent work has contested this term as it includes energy that is useful for the cells, for example motility and synthesis of bioimportant macromolecular compounds, along with energy that is wasted in "spillover" reactions and thus does not accurately represent the actual minimal energy required for life to survive. Following Hoehler and Jorgensen (2013), we elected to use the BPR, which is the energy flux required for the minimal amount of cellular functions to maintain a metabolically active cell.

Both the minimum maintenance energy and BPR are notoriously difficult to measure. Laboratory studies have yielded maintenance energies of order 10^{-5} kJ/(g·s) (Tijhuis *et al.*, 1993) to downwards of 10^{-7} kJ/(g·s) (Scholten and Conrad, 2000). The upper end of these values are especially problematic as it is ~ 26 times greater than the maintenance energy of a human body, which is not operating near its limits (Hoehler, 2004), and because it is temperature dependent.

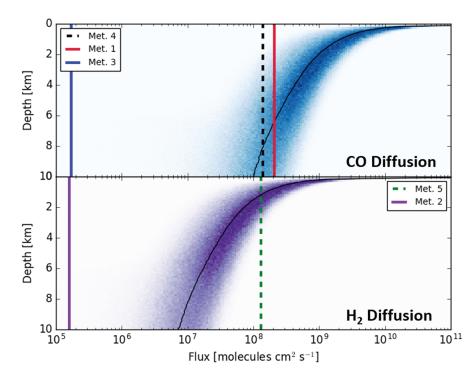


FIG. 3. Monte Carlo simulations calculating diffusive fluxes for CO and H₂ as a function of biotic layer depth. The resulting probability density is shown by colored bins. Solid black trendlines indicate median flux values, while vertical lines show maximum allowable biogenic sinks for each metabolism from our photochemical model calculations. The interception of these vertical lines with the median diffusion flux shows the maximum depth to which subsurface life could be exploiting atmospheric free energy. Below these depths, microbes would be limited by the downward diffusion of atmospheric gases through the regolith rather than the supply of reactants from photochemical reactions.

Minimum power requirements are especially difficult to characterize in natural settings (Van Bodegom, 2007; Onstott *et al.*, 2014; Kempes *et al.*, 2017), and recent work has shown that the BPR is a function of cellular volume (Kempes *et al.*, 2016). Measurements of sulfate-reducing bacteria in anoxic

marine sediments show BPRs as low as the aforementioned 3×10^{-23} kJ/(s·cell) (Lever *et al.*, 2015). This lower value agrees well with other recorded lower bounds on BPR of $\sim 5 \times 10^{-23}$ kJ/(s·cell) (Marschall *et al.*, 2010; LaRowe and Amend, 2015). A back-of-the-envelope attempt by LaRowe

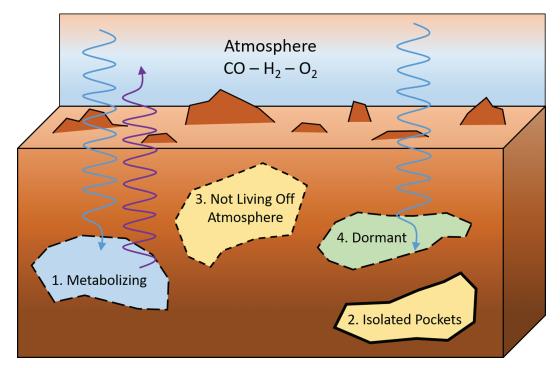


FIG. 4. Potential sites for life on Mars. This study estimates the maximum number of actively metabolizing microbes that are in communication with the atmosphere (1). There could be actively metabolizing microbes in small communities that are sealed off from the atmosphere (2) or are neither taking advantage of the available atmospheric free energy nor producing detectable byproducts (3). The number of dormant microbes (*e.g.* endospores) (4) could be indefinitely large but would not be evolutionarily advantageous for long-term survival on Mars. Downward arrows indicate fluxes of available free-energy reactants and upward arrows indicate flux of metabolized products into the atmosphere. All are assumed to be in contact with some form of liquid water (*e.g.* rare briny fluid pockets or a deep aquifer).

and Amend (2015) at arriving to a theoretical lower limit provides a BPR of $\sim 1\times 10^{-24}$ kJ/(s·cell) to prevent racemization of amino acids and thus population decay. However, the authors do note that this may not be a true limit and could be off by orders of magnitude. In addition, a BPR of $\sim 2\times 10^{-22}$ kJ/(s·cell) is more characteristic of microbes in these ultra-low energy environments rather than the lowest cell-specific BPR of $\sim 3\times 10^{-23}$ kJ/(s·cell). We have also been assuming in our calculations that the microbes are living at their lower limits with no population growth, with very long turnover rates, which may very well not be the case.

It is also important to note that this is a rigorous biomass estimate for actively metabolizing microbes in communication with the atmosphere. There are other possible scenarios for extant life on Mars (Fig. 4). For example, there could be an indefinitely large community of dormant microbes. Bacterial endospore states are highly durable to withstanding inclement environmental conditions and very low energy fluxes. Cellular turnover times of $\sim 10^3$ years have been found in deep biospheres on Earth (Lomstein et al., 2012). but the energy costs required of these dormant cells is unclear (Hoehler and Jorgensen, 2013). Nevertheless, it is unlikely that spore formation is evolutionarily advantageous for longterm survival in environments subject to harsh and degrading conditions, such as on Mars, due to the deterioration of DNA over time if molecular repair does not meet or exceed molecular damage (Mckay, 1997; Johnson et al., 2007; Cockell et al., 2016; Teodoro et al., 2018).

Additional biomass could also be concentrated in self-sustaining isolated pocket communities that are closed off from the atmosphere by impermeable lithological units (e.g., terrestrial microbes in Lin et al., 2006). However, these smaller isolated pockets would be harder to detect even with advanced drilling. One could also conceive of rarer unusual metabolisms that are in communication with the atmosphere but live off the regolith without excreting detectable products into the atmosphere. However, these ecosystems seem unlikely, given the abundance of free energy available in the atmosphere and the simplicity and primitive nature of the CO-reaction genes in terrestrial microbes.

Finally, the results from our diffusion calculations place additional restrictions on the abundance and distribution of life. We find that CO and H₂ can diffuse into the subsurface at much higher rates than our maximum calculated biological sinks in the upper 500 m. This suggests that life is not taking full advantage of this available free energy in the upper regolith as the photochemical energy far outweighs alternative subsurface sources (Jakosky and Shock, 1998). Therefore, any life in the upper crust is not limited by the available free energy but by some other factors (e.g., lack of liquid water or temperature dependency). Of course, this is consistent with the calculated instability of liquid water in the upper regolith, likely making it uninhabitable (Clifford et al., 2010). Future missions could also measure the CO and H2 fluxes at the surface of Mars to place tighter constraints on the availability of reactants into the subsurface.

It should also be noted that we consider these diffusion calculations to apply to the low to mid-latitudes as the high latitudes will have an ice-saturated cryosphere, which is not considered in this study. The lower latitudes will be desiccated due to the instability of ground ice with respect to the water vapor content of the atmosphere (Feldman *et al.*,

2004; Clifford *et al.*, 2010; Dundas *et al.*, 2014). Our calculations are relevant to the low to mid-latitudes because subsurface aquifers could be stable there over geological timescales (Grimm and Painter, 2009; Grimm *et al.*, 2017). As subsurface life presumably requires a liquid groundwater table, which is typically estimated around ≥ 5 km depth (Clifford *et al.*, 2010), the potential diffusive flux may be smaller than our calculated maximum biogenic surface sinks at these depths and further limit the maximum allowable biomass unless a minimum water activity level for life is maintained by a minimum H_2O layer thickness in the pore space somehow. In addition, we note our use of a 1D photochemical model assumes a globally distributed biosphere, but if life exists, it could be more spatially limited, so the biomass estimate would presumably be even lower.

5. Conclusions

The martian atmosphere has an untapped free energy source of $\sim 136 \text{ J/mol}$ predominantly contributed by the CO-O₂ ($\sim 133 \text{ J/mol}$) and H₂-O₂ ($\sim 3 \text{ J/mol}$) redox pairs (Krissansen-Totton *et al.*, 2016). This constant photochemically produced disequilibrium is far larger than any other energy source known to exist on Mars, such as internal heat. Given life's tendency to exploit chemical free energy, we show how these relatively high concentrations of CO and H₂ coexisting with high concentrations of O₂ can represent antibiosignatures and that any extant life on Mars is severely limited.

We quantify how much life could be taking advantage of this "free lunch" by using a 1D photochemical code and modeling biological activity as a downward surface flux of CO, and ramping it up until the modeled atmosphere diverges from observations. We find maximum feasible downward biogenic sinks of 1.5×10^8 and 1.9×10^8 molecules/(cm²·s) for CO and H₂, respectively, which are robust to uncertainties in observations and tunable model parameters. By using very conservative estimates on the minimal power requirements for microbial life and metabolic efficiencies, this equates to an upper limit of $\sim 10^{27}$ cells or the equivalent of ~ 1 million blue whales worth of metabolizing biomass that could be living off this atmospheric energy. This biomass estimate is highly dependent upon choice of cellular BPRs and typical microbes have higher requirements, thus any life is likely to be much smaller in extent. Diffusion calculations also imply that this biomass limit applies to life actively metabolizing within a few kilometers of the surface with the overall desiccation of the upper crust further limiting the possible biomass.

These results imply that any extant life on Mars is extremely limited in abundance. CO and $\rm H_2$ metabolisms are simple and phylogenetically widespread on Earth so that any microbial-like life on Mars should evolve to exploit this energy. In addition, more plausible assumptions on metabolic efficiencies, combined with recognizing the maximum gas diffusion limit into the crust realistically only reach small pockets of liquid water, imply that any biomass may be orders of magnitude smaller than presented in this study. Thus, what we present in this study is a robust upper limit on extant actively metabolizing martian life in contact with the atmosphere.

Also, the concept of using sufficient atmospheric CO and H_2 concentrations as an antibiosignature could be applied to future research in exoplanet characterization. While the

possibility for biogenic gases in exoplanet atmospheres as possible biosignatures has garnered much attention (*e.g.*, Seager and Bains, 2015; Schwieterman *et al.*, 2017), the possibility of antibiosignatures has received less (Wang *et al.*, 2016; Catling *et al.*, 2018). Where sufficient knowledge is known on the atmospheric composition, the presence of high concentrations of CO or H₂ in thermodynamic disequilibrium would allow for a deduction of an absence of a productive biosphere.

Acknowledgments

We thank Drew Gorman-Lewis, Adam Stevens, and Kevin Zahnle for helpful discussions. We also thank Jim Kasting, Tullis Onstott, and an anonymous reviewer, whose comments greatly improved the rigor and clarity of the article. This work was supported by NASA Astrobiology Institute's Virtual Planetary Laboratory, grant NNA13AA 93A, and by NASA Exobiology Program, grant NNX15 AL23G, awarded to D.C.C. J.K.-T. is supported by NASA Headquarters under the NASA Earth and Space Science Fellowship program, grant NNX15AR63H. The article was completed while D.C.C. was a Leverhulme Trust Visiting Professor at the University of Cambridge, United Kingdom.

Author Disclosure Statement

No competing financial interests exist.

References

- Bakken, L.R. (1985) Separation and purification of bacteria from soil. *Appl Environ Microbiol* 49:1482–1487.
- Bar-On, Y.M., Phillips R., and Milo, R. (2018) The biomass distribution on Earth. *Proc Natl Acad Sci U S A* 115:6506–6511.
- Bradley, J.A., Amend, J.P., and LaRowe, D.E. (2018) Bioenergetic controls on microbial ecophysiology in marine sediments. Front Microbiol 9:180.
- Bratbak, G. and Dundas, I. (1984). Bacterial dry matter content and biomass estimations. Appl Environ Microbiol 48:755–757.
- Catling, D.C. and Kasting, J.F. (2017) *Atmospheric Evolution on Inhabited and Lifeless Worlds*. Cambridge University Press. Cambridge, UK.
- Catling, D.C., Krissansen-Totton, J., Kiang, N.Y., Crisp, D., Robinson, T.D., DasSarma, S., Rushby, A., Del Genio, A., Bains, W., and Domagal-Goldman, S. (2018) Exoplanet biosignatures: a framework for their assessment. *Astrobiology* 18:709–738.
- Clark, B.C., Baird, A., Rose, H.J., Toulmin, P., Keil, K., Castro, A.J., Kelliher, W.C., Rowe, C.D., and Evans, P.H. (1976) Inorganic analyses of Martian surface samples at the Viking landing sites. *Science* 194:1283–1288.
- Clifford, S.M. (1993) A model for the hydrologic and climatic behavior of water on Mars. *J Geophys Res Planets* 98:10973–11016.
- Clifford, S.M., Lasue, J., Heggy, E., Boisson, J., McGovern, P., and Max, M.D. (2010) Depth of the Martian cryosphere: revised estimates and implications for the existence and detection of subpermafrost groundwater. *J Geophys Res Planets* 115:E07001.
- Cockell, C.S., Catling, D.C., Davis, W.L., Snook, K., Kepner, R.L., Lee, P., and McKay, C.P. (2000) The ultraviolet environment of Mars: biological implications past, present, and future. Icarus 146:343–359.
- Cockell, C.S., Bush, T., Bryce, C., Direito, S., Fox-Powell, M., Harrison, J., Lammer, H., Landenmark, H., Martin-Torres, J.,

and Nicholson, N. (2016) Habitability: a review. *Astrobiology* 16:89–117.

- Dartnell, L.R., Desorgher, L., Ward, J., and Coates, A. (2007) Modelling the surface and subsurface martian radiation environment: implications for astrobiology. *Geophys Res Lett* 34: L02207.
- Dundas, C.M., Byrne, S., McEwen, A.S., Mellon, M.T., Kennedy, M.R., Daubar, I.J., and Saper, L. (2014) HiRISE observations of new impact craters exposing Martian ground ice. *J Geophys Res Planets* 119:109–127.
- Ehlmann, B., Mustard, J., and Murchie, S. (2010) Geologic setting of serpentine deposits on Mars. *Geophys Res Lett* 37:L06201.
- Falkowski, P.G., Fenchel, T., and Delong, E.F. (2008) The microbial engines that drive Earth's biogeochemical cycles. *Science* 320:1034–1039.
- Feldman, W.C., Prettyman, T.H., Maurice, S., Plaut, J., Bish, D., Vaniman, D., Mellon, M., Metzger, A., Squyres, S., and Karunatillake, S. (2004) Global distribution of near-surface hydrogen on Mars. *J Geophys Res Planets* 109:E09006.
- Ferry, J.G. and House, C.H. (2005) The stepwise evolution of early life driven by energy conservation. *Mol Biol Evol* 23:1286–1292.
- Formisano, V., Atreya, S., Encrenaz, T., Ignatiev, N., and Giuranna, M. (2004) Detection of methane in the atmosphere of Mars. *Science* 306:1758–1761.
- Franz, H.B., Trainer, M.G., Malespin, C.A., Mahaffy, P.R., Atreya, S.K., Becker, R.H., Benna, M., Conrad, P.G., Eigenbrode, J.L., and Freissinet, C. (2017) Initial SAM calibration gas experiments on Mars: quadrupole mass spectrometer results and implications. *Planet Space Sci* 138:44–54.
- Grimm, R.E. and Painter, S.L. (2009) On the secular evolution of groundwater on Mars. *Geophys Res Lett* 36.
- Grimm, R.E., Harrison, K.P., Stillman, D.E., and Kirchoff, M.R. (2017) On the secular retention of ground water and ice on Mars. J Geophys Res Planets 122:94–109.
- Haberle, R.M. (2013) Estimating the power of Mars' greenhouse effect. *Icarus* 223:619–620.
- Haberle, R.M., Forget, F., Colaprete, A., Schaeffer, J., Boynton, W.V., Kelly, N.J., and Chamberlain, M.A. (2008) The effect of ground ice on the Martian seasonal CO₂ cycle. *Planet. Space Sci* 56:251–255.
- Hanna, J.C. and Phillips, R.J. (2005) Hydrological modeling of the Martian crust with application to the pressurization of aquifers. J Geophys Res Planets 110:E01004.
- Hitchcock, D.R. and Lovelock, J.E. (1967) Life detection by atmospheric analysis. *Icarus* 7:149–159.
- Hoehler, T. (2004) Biological energy requirements as quantitative boundary conditions for life in the subsurface. *Geobiology* 2:205–215.
- Hoehler, T.M. and Jorgensen, B.B. (2013) Microbial life under extreme energy limitation. *Nat Rev Microbiol* 11:83–94.
- Huber, C. and Wächtershäuser, G. (1997) Activated acetic acid by carbon fixation on (Fe, Ni) S under primordial conditions. *Science* 276:245–247.
- Jakosky, B.M. and Shock, E.L. (1998) The biological potential of Mars, the early Earth, and Europa. J Geophys Res Planets 103:19359–19364.
- Jakosky, B., Brain, D., Chaffin, M., Curry, S., Deighan, J., Grebowsky, J., Halekas, J., Leblanc, F., Lillis, R., and Luhmann, J. (2018) Loss of the Martian atmosphere to space: present-day loss rates determined from MAVEN observations and integrated loss through time. *Icarus*. 315:146–157
- Jeoung, J.-H. and Dobbek, H. (2007) Carbon dioxide activation at the Ni, Fe-cluster of anaerobic carbon monoxide dehydrogenase. *Science* 318:1461–1464.

- Ji, M., Greening, C., Vanwonterghem, I., Carere, C.R., Bay, S.K., Steen, J.A., Montgomery, K., Lines, T., Beardall, J., and van Dorst, J. (2017) Atmospheric trace gases support primary production in Antarctic desert surface soil. *Nature* 552:400.
- Johnson, S.S., Hebsgaard, M.B., Christensen, T.R., Mastepanov, M., Nielsen, R., Munch, K., Brand, T., Gilbert, M.T.P., Zuber, M.T., and Bunce, M. (2007) Ancient bacteria show evidence of DNA repair. *Proc Natl Acad Sci U S A* 104:14401–14405.
- Kallmeyer, J., Pockalny, R., Adhikari, R.R., Smith, D.C., and D'Hondt, S. (2012) Global distribution of microbial abundance and biomass in subseafloor sediment. *Proc Natl Acad Sci U S A* 109:16213–16216.
- Kasting, J.F. (1979) Evolution of oxygen and ozone in the Earth's atmosphere. PhD Thesis. University of Michigan, p. 259.
- Kempes, C.P., Wang, L., Amend, J.P., Doyle, J., and Hoehler, T. (2016) Evolutionary tradeoffs in cellular composition across diverse bacteria. *ISME J* 10:2145.
- Kempes, C.P., van Bodegom, P.M., Wolpert, D., Libby, E., Amend, J., and Hoehler, T. (2017) Drivers of bacterial maintenance and minimal energy requirements. *Front Microbiol* 31.
- Klein, H.P. (1978) The Viking biological experiments on Mars. *Icarus* 34:666–674.
- Klein, H.P. (1998) The search for life on Mars: what we learned from Viking. *J Geophys Res Planets* 103:28463–28466.
- Krasnopolsky, V. (1993) Photochemistry of the Martian atmosphere (mean conditions). *Icarus* 101:313–332.
- Krasnopolsky, V.A. and Feldman, P.D. (2001) Detection of molecular hydrogen in the atmosphere of Mars. *Science* 294:1914–1917.
- Krasnopolsky, V.A., Maillard, J.P., and Owen, T.C. (2004) Detection of methane in the martian atmosphere: evidence for life? *Icarus* 172:537–547.
- Krissansen-Totton, J., Bergsman, D.S., and Catling, D.C. (2016) On detecting biospheres from chemical thermodynamic disequilibrium in planetary atmospheres. *Astrobiology* 16:39–67.
- Lammer, H., Lichtenegger, H., Kolb, C., Ribas, I., Guinan, E., Abart, R., and Bauer, S. (2003) Loss of water from Mars: implications for the oxidation of the soil. *Icarus* 165:9–25.
- LaRowe, D.E. and Amend, J.P. (2015) Power limits for microbial life. Front Microbiol 6:718.
- Lessner, D.J., Li, L., Li, Q., Rejtar, T., Andreev, V.P., Reichlen, M., Hill, K., Moran, J.J., Karger, B.L., and Ferry, J.G. (2006) An unconventional pathway for reduction of CO₂ to methane in CO-grown *Methanosarcina acetivorans* revealed by proteomics. *Proc Natl Acad Sci U S A* 103:17921–17926.
- Lever, M.A., Rogers, K.L., Lloyd, K.G., Overmann, J., Schink, B., Thauer, R.K., Hoehler, T.M., and Jørgensen, B.B. (2015) Life under extreme energy limitation: a synthesis of laboratory-and field-based investigations. *FEMS Microbiol Rev* 39:688–728.
- Lillis, R.J., Deighan, J., Fox, J.L., Bougher, S.W., Lee, Y., Combi, M.R., Cravens, T.E., Rahmati, A., Mahaffy, P.R., and Benna, M. (2017) Photochemical escape of oxygen from Mars: first results from MAVEN in situ data. *J Geophys Res* Space Phys 122:3815–3836.
- Lin, L.-H., Wang, P.-L., Rumble, D., Lippmann-Pipke, J., Boice, E., Pratt, L.M., Lollar, B.S., Brodie, E.L., Hazen, T.C., and Andersen, G.L. (2006) Long-term sustainability of a highenergy, low-diversity crustal biome. *Science* 314:479–482.
- Lomstein, B.A., Langerhuus, A.T., D'Hondt, S., Jørgensen, B.B., and Spivack, A.J. (2012) Endospore abundance, microbial growth and necromass turnover in deep sub-seafloor sediment. *Nature* 484:101.
- Lovelock, J.E. (1965) A physical basis for life detection experiments. *Nature* 207:568–570.

- Lovelock, J.E. (1975) Thermodynamics and the recognition of alien biospheres. *Proc R Soc Lond B Biol Sci* 189:167–181.
- Lovelock, J.E. (1979) *Gaia: A New Look at Life on Earth.*Oxford University Press, Oxford.
- Lovelock, J. (1988) *The Ages of Gaia*. W.W. Norton, New York.
 Mahaffy, P. R., Webster, C. R., Atreya, S. K., Franz, H., Wong,
 M., Conrad, P. G., et al. (2013). Abundance and Isotopic Composition of Gases in the Martian Atmosphere from the Curiosity Rover. *Science* 341:263–266.
- Marschall, E., Jogler, M., Henßge, U., and Overmann, J. (2010) Large-scale distribution and activity patterns of an extremely low-light-adapted population of green sulfur bacteria in the Black Sea. *Environ Microbiol* 12:1348–1362.
- Martín-Torres, F.J., Zorzano, M.-P., Valentín-Serrano, P., Harri, A.-M., Genzer, M., Kemppinen, O., Rivera-Valentin, E.G., Jun, I., Wray, J., and Madsen, M.B. (2015) Transient liquid water and water activity at Gale crater on Mars. *Nat Geosci* 8:357–361.
- McKay, C.P. (1997) The search for life on Mars. In *Planetary* and *Interstellar Processes Relevant to the Origins of Life*, edited by D.C.B. Whittet, Springer, Dordrecht, NL, pp. 263–289.
- McMahon, S., Parnell, J., and Blamey, N.J. (2016) Evidence for seismogenic hydrogen gas, a potential microbial energy source on Earth and Mars. *Astrobiology* 16:690–702.
- Mellon, M.T., Jakosky, B.M., and Postawko, S.E. (1997) The persistence of equatorial ground ice on Mars. *J Geophys Res Planets* 102:19357–19369.
- Meyer, O. and Schlegel, H.G. (1983) Biology of aerobic carbon monoxide-oxidizing bacteria. *Annu Rev Microbiol* 37:277–310.
- Michalski, J.R., Cuadros, J., Niles, P.B., Parnell, J., Rogers, A.D., and Wright, S.P. (2013) Groundwater activity on Mars and implications for a deep biosphere. *Nat Geosci* 6:133.
- Mickol, R. and Kral, T. (2017) Low pressure tolerance by methanogens in an aqueous environment: implications for subsurface life on Mars. *Orig Life Evol Biosph* 47:511–532.
- Mumma, M.J., Villanueva, G.L., Novak, R.E., Hewagama, T., Bonev, B.P., DiSanti, M.A., Mandell, A.M., and Smith, M.D. (2009) Strong release of methane on Mars in northern summer 2003. *Science* 323:1041–1045.
- Myers, M.R. and King, G.M. (2017) Perchlorate-coupled carbon monoxide (CO) oxidation: evidence for a plausible microbe-mediated reaction in Martian brines. *Front Microbiol* 8:2571.
- Nair, H., Allen, M., Anbar, A.D., Yung, Y.L., and Clancy, R.T. (1994) A photochemical model of the martian atmosphere. *Icarus* 111:124–150.
- Ojha, L., Wilhelm, M.B., Murchie, S.L., McEwen, A.S., Wray, J.J., Hanley, J., Massé, M., and Chojnacki, M. (2015) Spectral evidence for hydrated salts in recurring slope lineae on Mars. *Nat Geosci* 8:829–832.
- Onstott, T., McGown, D., Kessler, J., Lollar, B.S., Lehmann, K., and Clifford, S. (2006) Martian CH₄: sources, flux, and detection. *Astrobiology* 6:377–395.
- Onstott, T., Magnabosco, C., Aubrey, A., Burton, A., Dworkin, J., Elsila, J., Grunsfeld, S., Cao, B., Hein, J., and Glavin, D. (2014) Does aspartic acid racemization constrain the depth limit of the subsurface biosphere? *Geobiology* 12:1–19.
- Orosei, R., Lauro, S., Pettinelli, E., Cicchetti, A., Coradini, M., Cosciotti, B., Di Paolo, F., Flamini, E., Mattei, E., and Pajola, M. (2018) Radar evidence of subglacial liquid water on Mars. *Science* 361:490–493.
- Pavlov, A., Vasilyev, G., Ostryakov, V., Pavlov, A., and Mahaffy, P. (2012) Degradation of the organic molecules in the shallow subsurface of Mars due to irradiation by cosmic rays. *Geophys Res Lett* 39:L13202.

Ragsdale, S.W. (2004) Life with carbon monoxide. Crit Rev Biochem Mol Biol 39:165–195.

- Rennó, N.O., Bos, B.J., Catling, D., Clark, B.C., Drube, L., Fisher, D., Goetz, W., Hviid, S.F., Keller, H.U., and Kok, J.F. (2009) Possible physical and thermodynamical evidence for liquid water at the Phoenix landing site. *J Geophys Res Planets* 114:E00E03.
- Rother, M. and Metcalf, W.W. (2004) Anaerobic growth of *Methanosarcina acetivorans* C2A on carbon monoxide: an unusual way of life for a methanogenic archaeon. *Proc Natl Acad Sci U S A* 101:16929–16934.
- Rummel, J.D., Beaty, D.W., Jones, M.A., Bakermans, C., Barlow, N.G., Boston, P.J., Chevrier, V.F., Clark, B.C., de Vera, J.-P.P., and Gough, R.V. (2014) A new analysis of Mars "special regions": findings of the second MEPAG Special Regions Science Analysis Group (SR-SAG2). Astrobiology 14:887–968.
- Scholten, J.C. and Conrad, R. (2000) Energetics of syntrophic propionate oxidation in defined batch and chemostat cocultures. *Appl Environ Microbiol* 66:2934–2942.
- Schwieterman, E.W., Kiang, N.Y., Parenteau, M.N., Harman, C.E., DasSarma, S., Fisher, T.M., Arney, G.N., Hartnett, H.E., Reinhard, C.T., and Olson, S.L. (2017) Exoplanet biosignatures: a review of remotely detectable signs of life. arXiv:1705.05791.
- Seager, S. and Bains, W. (2015) The search for signs of life on exoplanets at the interface of chemistry and planetary science. Sci Adv 1:e1500047.
- Seinfeld, J.H. and Pandis, S.N. (2016) Atmospheric Chemistry and Physics: From Air Pollution to Climate Change. John Wiley & Sons. Hoboken, N.J.
- Sholes, S.F., Smith, M.L., Claire, M.W., Zahnle, K.J., and Catling, D.C. (2017) Anoxic atmospheres on Mars driven by volcanism: implications for past environments and life. *Icarus* 290:46–62.
- Sizemore, H.G. and Mellon, M.T. (2008) Laboratory characterization of the structural properties controlling dynamical gas transport in Mars-analog soils. *Icarus* 197:606–620.
- Smith, M.L., Claire, M.W., Catling, D.C., and Zahnle, K.J. (2014) The formation of sulfate, nitrate and perchlorate salts in the martian atmosphere. *Icarus* 231:51–64.
- Stevens, A.H., Patel, M.R., and Lewis, S.R. (2015) Numerical modelling of the transport of trace gases including methane in the subsurface of Mars. *Icarus* 250:587–594.
- Stevens, A.H., Patel, M.R., and Lewis, S.R. (2017) Modelled isotopic fractionation and transient diffusive release of methane from potential subsurface sources on Mars. *Icarus* 281: 240–247.
- Techtmann, S.M., Colman, A.S., and Robb, F.T. (2009) "That which does not kill us only makes us stronger": the role of carbon monoxide in thermophilic microbial consortia. *Environ Microbiol* 11:1027–1037.
- Teodoro, L., Davila, A., Elphic, R.C., Hamilton, D., McKay, C., and Quinn, R. (2018) Habitability and biomarker preservation in the martian near-surface radiation environment. In *From Habitability to Life on Mars*, edited by N.A. Cabrol and E.A. Grin, Elsevier, Cambridge, MA, pp 211–231.
- Tijhuis, L., Van Loosdrecht, M.C., and Heijnen, J. (1993) A thermodynamically based correlation for maintenance Gibbs energy requirements in aerobic and anaerobic chemotrophic growth. *Biotechnol Bioeng* 42:509–519.
- Toner, J.D., Catling, D.C., and Light, B. (2014) The formation of supercooled brines, viscous liquids, and low-temperature perchlorate glasses in aqueous solutions relevant to Mars. *Icarus* 233:36–47.
- Ueno, Y., Yamada, K., Yoshida, N., Maruyama, S., and Isozaki, Y. (2006) Evidence from fluid inclusions for microbial methanogenesis in the early Archaean era. *Nature* 440:516.

Van Bodegom, P. (2007) Microbial maintenance: a critical review on its quantification. *Microb Ecol* 53:513–523.

- Wadsworth, J. and Cockell, C.S. (2017) Perchlorates on Mars enhance the bacteriocidal effects of UV light. *Sci Rep* 7:4662.
- Wang, Y., Tian, F., Li, T., and Hu, Y. (2016) On the detection of carbon monoxide as an anti-biosignature in exoplanetary atmospheres. *Icarus* 266:15–23.
- Webster, C.R., Mahaffy, P.R., Atreya, S.K., Flesch, G.J., Mischna, M.A., Meslin, P.-Y., Farley, K.A., Conrad, P.G., Christensen, L.E., Pavlov, A.A., Martín-Torres, J., Zorzano, M.P., McConnochie, T.H., Owen, T., Eigenbrode, J.L., Glavin, D.P., Steele, A., Malespin, C.A., Archer, P.D. Jr., Sutter, B., Coll, P., Freissinet, C., McKay, C.P., Moores, J.E., Schwenzer, S.P., Bridges, J.C., Navarro-Gonzalez, R., Gellert, R., and Lemmon, M.T.; MSL Science Team. (2015) Mars methane detection and variability at Gale crater. Science 347:415–417.
- Webster, C.R., Mahaffy, P.R., Atreya, S.K., Moores, J.E., Flesch, G.J., Malespin, C., McKay, C.P., Martinez, G., Smith, C.L., and Martin-Torres, J. (2018) Background levels of methane in Mars' atmosphere show strong seasonal variations. *Science* 360:1093–1096.
- Weiss, B.P., Yung, Y.L., and Nealson, K.H. (2000) Atmospheric energy for subsurface life on Mars? *Proc Natl Acad Sci U S A* 97:1395–1399.
- Weiss, M.C., Sousa, F.L., Mrnjavac, N., Neukirchen, S., Roettger, M., Nelson-Sathi, S., and Martin, W.F. (2016) The physiology and habitat of the last universal common ancestor. *Nat Microbiol* 1:16116.
- Wolfe, J.M. and Fournier, G.P. (2018) Horizontal gene transfer constrains the timing of methanogen evolution. *Nat Ecol Evol* 2:897.
- Yung, Y.L., Chen, P., Nealson, K., Atreya, S., Beckett, P., Blank, J.G., Ehlmann, B., Eiler, J., Etiope, G., and Ferry, J.G. (2018) Methane on Mars and habitability: challenges and responses. *Astrobiology* 18:1221–1242.
- Zahnle, K. (2015) Play it again, SAM. Science 347:370–371.
 Zahnle, K., Haberle, R.M., Catling, D.C., and Kasting, J.F. (2008) Photochemical instability of the ancient Martian atmosphere. J Geophys Res Planets 113:E11004.
- Zahnle, K., Freedman, R.S., and Catling, D.C. (2011) Is there methane on Mars? *Icarus* 212:493–503.

Address correspondence to:
Steven F. Sholes
Department of Earth and Space Sciences
University of Washington
Johnson Hall 070, Box 351310
4000 15th Avenue NE
Seattle, WA 98195-1310

E-mail: sfsholes@uw.edu

Submitted 12 February 2018 Accepted 17 November 2018

Abbreviations Used

1D = one-dimensional

BPR = basal power requirement

CODH = carbon monoxide dehydrogenase

MAVEN = Mars Atmosphere and Volatile

EvolutioN

Met. = metabolism